

Package ‘peakPantheR’

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Title Peak Picking and Annotation of High Resolution Experiments

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Description An automated pipeline for the detection, integration and reporting of predefined features across a large number of mass spectrometry data files.
It enables the real time annotation of multiple compounds in a single file, or the parallel annotation of multiple compounds in multiple files.
A graphical user interface as well as command line functions will assist in assessing the quality of annotation and update fitting parameters until a satisfactory result is obtained.

Depends R (>= 4.2)

Imports foreach (>= 1.4.4), doParallel (>= 1.0.11), ggplot2 (>= 3.5.0), gridExtra (>= 2.3), MSnbase (>= 2.4.0), mzR (>= 2.12.0), stringr (>= 1.2.0), methods (>= 3.4.0), XML (>= 3.98.1.10), minpack.lm (>= 1.2.1), scales (>= 0.5.0), shiny (>= 1.0.5), bslib, shinycssloaders (>= 1.0.0), DT (>= 0.15), pracma (>= 2.2.3), utils, lubridate, svglite (>= 2.1.1)

biocViews MassSpectrometry, Metabolomics, PeakDetection

License GPL-3

BugReports <https://github.com/phenomecentre/peakPantheR/issues/new>

URL <https://github.com/phenomecentre/peakPantheR>

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acquisitionTime,peakPantheRAnnotation-method

acquisitionTime accessor returns value as *POSIXct*

Description

acquisitionTime accessor returns value as POSIXct

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
acquisitionTime(object)
```

Arguments

object peakPantheRAnnotation

Value

(POSIXct) A character vector of acquisition date-time (converted from POSIXct) or NA

Examples

```
if(requireNamespace('faahKO')){
  ## Initialise a peakPantherAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahKO)
  spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

  # targetFeatTable
  targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
    c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
  targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
    522.2, 522.205222)
  targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
    496.2, 496.204962)
  targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
    FUN.VALUE=numeric(2))

  annotation <- peakPantherAnnotation(spectraPaths=spectraPaths,
    targetFeatTable=targetFeatTable)

  ## acquisitionTime can only be extracted from NetCDF files
  acquisitionTime(annotation)
  # [1] NA NA NA
}
```

annotationDiagnosticMultiplot

Generate a multiplot of all diagnostic plots

Description

Generate a multiplot of all diagnostic plots (as generated by annotationDiagnosticPlots()) for each compound

Usage

```
annotationDiagnosticMultiplot(annotationDiagnosticPlotList)
```

Arguments

```
annotationDiagnosticPlotList
```

(list) List of (one per compound) of list of diagnostic plots as generated by annotationDiagnosticPlots()

Value

A list of multiplots (one per compound)

annotationDiagnosticPlots,peakPantheRAnnotation-method
Generate fit diagnostic plots

Description

Generate fit diagnostic plots for each ROI: EICFit the raw data and detected feature fit, rtPeakwidthVert detected peaks retention time apex and peakwidth (vertical and no run order), rtPeakwidthHorzRunOrder detected peaks retention time apex and peakwidth by run order, mzPeakwidthHorzRunOrder detected peaks m/z apex and peakwidth by run order, areaRunOrder detected peaks area by run order, rtHistogram histogram of detected peaks retention time, mzHistogram histogram of detected peaks m/z, areaHistogram histogram of detected peaks area.

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
annotationDiagnosticPlots(
  object,
  sampleColour,
  sampling,
  verbose
)
```

Arguments

object	(peakPantheRAnnotation) Annotated peakPantheRAnnotation object
sampleColour	(str) NULL or vector colour for each sample
sampling	(int) Number of points to employ when plotting fittedCurve
verbose	(bool) if TRUE message the plot generation progress

Value

A list (one list per compound) of diagnostic plots: result[[i]]\$EICFit, result[[i]]\$rtPeakwidthVert, result[[i]]\$rtPeakwidthHorzRunOrder, result[[i]]\$mzPeakwidthHorzRunOrder, result[[i]]\$areaRunOrder, result[[i]]\$rtHistogram, result[[i]]\$mzHistogram, result[[i]]\$areaHistogram, result[[i]]\$title

Examples

```
if(requireNamespace('faahKO')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahKO)
  spectraPaths <- c(system.file('cdf/K0/ko15.CDF', package = 'faahKO'),
                    system.file('cdf/K0/ko16.CDF', package = 'faahKO'),
                    system.file('cdf/K0/ko18.CDF', package = 'faahKO'))
```

```

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

annotationDiagnosticPlots(emptyAnnotation)
# Warning: the object has not been annotated, return an empty diagnostic plot
# list
# [[1]]
# NULL
# [[2]]
# NULL
}

```

annotationParamsDiagnostic,peakPantheRAnnotation-method

Set uROI and FIR based on annotation results

Description

Set updated ROI (uROI) and Fallback Integration Regions (FIR) based on the annotation results. If the object is not annotated, it is returned untouched. ROI is not modified. If uROI exist they are left untouched, otherwise they are set as the minimum and maximum found peaks limits (+/-5% of ROI in retention time). If FIR are used they are left untouched, otherwise they are set as the median of the found limits (rtMin, rtMax, mzMin, mzMax).

Usage

```

## S4 method for signature 'peakPantheRAnnotation'
annotationParamsDiagnostic(
  object,
  verbose
)

```

Arguments

object (peakPantheRAnnotation) Annotated peakPantheRAnnotation object
 verbose (bool) If TRUE message progress of uROI and FIR calculation

Value

(peakPantheRAnnotation) object with updated ROI and FIR set from annotation results

Examples

```

if(requireNamespace('faahKO')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahKO)
  spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

  # targetFeatTable
  targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
    c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
  targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
    522.2, 522.205222)
  targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
    496.2, 496.204962)
  targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
    FUN.VALUE=numeric(2))

  emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
    targetFeatTable=targetFeatTable)

  annotationParamsDiagnostic(emptyAnnotation, verbose=TRUE)
  # Warning: the object has not been annotated, return the object untouched
  # An object of class peakPantheRAnnotation
  # 2 compounds in 3 samples.
  # updated ROI do not exist (uROI)
  # does not use updated ROI (uROI)
  # does not use fallback integration regions (FIR)
  # is not annotated
}

```

```

annotationTable, peakPantheRAnnotation-method
annotationTable accessor

```

Description

annotationTable returns a dataframe (row samples, col compounds) filled with a specific peakTable column

Usage

```

## S4 method for signature 'peakPantheRAnnotation'
annotationTable(object, column)

```

Arguments

object	peakPantheRAnnotation
column	a peakTable columns

Value

(data.frame) (row samples, col compounds) filled with a specific peakTable column

Examples

```
if(requireNamespace('faahKO')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahKO)
  spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

  # targetFeatTable
  targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
    c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
  targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
    522.2, 522.205222)
  targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
    496.2, 496.204962)
  targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
    FUN.VALUE=numeric(2))

  annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
    targetFeatTable=targetFeatTable)

  ## default values without annotation
  annotationTable(annotation)
  #
  # ID-1 ID-2
  # C:/R/R-3.6.0/library/faahKO/cdf/KO/ko15.CDF NA NA
  # C:/R/R-3.6.0/library/faahKO/cdf/KO/ko16.CDF NA NA
  # C:/R/R-3.6.0/library/faahKO/cdf/KO/ko18.CDF NA NA
}
```

annotation_diagnostic_multiplot_UI_helper

UI diagnostic plot helper - single feature multiplot

Description

Return a ggplot object of a feature diagnostic multiplot

Usage

```
annotation_diagnostic_multiplot_UI_helper(
  cpdNb,
  annotation,
  splNum = NULL,
  splColrColumn = NULL,
  ...
)
```


Arguments

cpdNb (int) position of the feature to extract (1 to nbCpd)
 annotation (peakPantheRAnnotation) Annotation object
 splNum (int) NULL or number of spectra to plot, chosen randomly from all spectra. If NULL or equal to the total number of spectra, plot all spectra
 splColrColumn (str) NULL, None or a spectraMetadata column for colouring each sample
 ... Additional parameters for plotting

Value

(ggplotObject) Diagnostic multiplot for a feature

Examples

```
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# Plot of an empty annotation
annotation_diagnostic_multiplot_UI_helper(cpdNb = 2,
  annotation = emptyAnnotation,
  splNum = NULL,
  splColrColumn = NULL)

# Warning: the object has not been annotated, return an empty diagnostic
# plot list
```

annotation_fit_summary_UI_helper

UI diagnostic table - fit summary

Description

Return a table of fit statistic (ratio of peaks found, ratio of peaks filled, ppm error, RT deviation)

Usage

```
annotation_fit_summary_UI_helper(annot)
```

Arguments

annot (peakPantheRAnnotation) Annotation object

Value

(data.frame) Fit statistics

Examples

```
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# statistics of an empty annotation
annotation_fit_summary_UI_helper(emptyAnnotation)
#           Ratio peaks found (%) Ratio peaks filled (%) ppm error
# ID-1 - Cpd 1                NA                0                NaN
# ID-2 - Cpd 2                NA                0                NaN
#           RT deviation (s)
# ID-1 - Cpd 1                NaN
# ID-2 - Cpd 2                NaN
```

```
annotation_showMethod_UI_helper
```

UI show annotation helper - list of properties

Description

show method specific to the UI, that returns each field in a named list to ease display

Usage

```
annotation_showMethod_UI_helper(annotation)
```

Arguments

annotation (peakPantherAnnotation) Object to describe

Value

(list) Named list of annotation properties

Examples

```
# Initialise an empty annotation, no uROI, no use of FIR
annotInit <- peakPantheRAnnotation()

# return properties
annotation_showMethod_UI_helper(annotInit)
# $nbCompounds
# [1] 0
#
# $nbSamples
# [1] 0
#
# $uROIExist
# [1] FALSE
#
# $useUROI
# [1] FALSE
#
# $useFIR
# [1] FALSE
#
# $isAnnotated
# [1] FALSE
```

annotation_showText_UI_helper

UI show annotation helper - UI sidebar string

Description

Return a text description of an annotation for UI presentation

Usage

```
annotation_showText_UI_helper(annotProp)
```

Arguments

annotProp (list) Named list of annotation properties as created by annotation_showMethod_UI_helper()

Value

(str) Textual description of the annotation to show on UI

Examples

```
# Input
properties_default <- list(nbCompounds = 0,
                          nbSamples = 0,
                          uROIExist = FALSE,
                          useUROI = FALSE,
                          useFIR = FALSE,
                          isAnnotated = FALSE)

# Generate description
annotation_showText_UI_helper(properties_default)
# [[1]]
# [1] "Not annotated"
#
# [[2]]
# [1] "0 compounds"
#
# [[3]]
# [1] "0 samples"
#
# [[4]]
# [1] "updated ROI do not exist (uROI)"
#
# [[5]]
# [1] "does not use updated ROI (uROI)"
#
# [[6]]
# [1] "does not use fallback integration regions (FIR)"
```

cpdID,peakPantheRAnnotation-method
cpdID accessor

Description

cpdID accessor

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
cpdID(object)
```

Arguments

object peakPantheRAnnotation

Value

(str) A character vector of compound IDs, of length number of compounds

Examples

```

if(requireNamespace('faahKO')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahKO)
  spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

  # targetFeatTable
  targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
    c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin',
      'mz', 'mzMax'))), stringsAsFactors=FALSE)
  targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
    522.2, 522.205222)
  targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
    496.2, 496.204962)
  targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
    FUN.VALUE=numeric(2))

  annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
    targetFeatTable=targetFeatTable)

  cpdID(annotation)
  # [1] 'ID-1' 'ID-2'
}

```

cpdMetadata,peakPantheRAnnotation-method
cpdMetadata accessor

Description

cpdMetadata accessor

Usage

```

## S4 method for signature 'peakPantheRAnnotation'
cpdMetadata(object)

```

Arguments

object peakPantheRAnnotation

Value

(data.frame) A data.frame of compound metadata, with compounds as row and metadata as columns

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
                        c('cpdID','cpdName','rtMin','rt','rtMax','mzMin','mz',
                        'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                        522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                        496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
                                  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
                                   targetFeatTable=targetFeatTable)

## default values not initialised
cpdMetadata(annotation)
# data frame with 0 columns and 2 rows
}

```

cpdName,peakPantheRAnnotation-method
cpdName accessor

Description

cpdName accessor

Usage

```

## S4 method for signature 'peakPantheRAnnotation'
cpdName(object)

```

Arguments

object peakPantheRAnnotation

Value

(str) A character vector of compound names, of length number of compounds

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/K0/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/K0/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/K0/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
                          c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
                          'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                        522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                        496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
                                  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
                                   targetFeatTable=targetFeatTable)

cpdName(annotation)
# [1] 'Cpd 1' 'Cpd 2'
}

```

dataPoints,peakPantheRAnnotation-method
dataPoints accessor

Description

dataPoints accessor

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
dataPoints(object)
```

Arguments

object peakPantheRAnnotation

Value

A list of length number of spectra files. Each list element is a *ROI>DataPoint* list of data.frame of raw data points for each ROI/uROI (retention time 'rt', mass 'mz' and intensity 'int' (as column) of each raw data points (as row))

Value

(float) Extracted Ion Chromatogram aggregated across m/z in each scan

Examples

```

if(requireNamespace('faahK0')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahK0)
spectraPaths <- c(system.file('cdf/K0/ko15.CDF', package = 'faahK0'),
                  system.file('cdf/K0/ko16.CDF', package = 'faahK0'),
                  system.file('cdf/K0/ko18.CDF', package = 'faahK0'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
                        c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
                          'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                        522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                        496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
                                   FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
                                   targetFeatTable=targetFeatTable)

## default values without annotation
EICs(annotation)
# [[1]]
# list()
# [[2]]
# list()
# [[3]]
# list()
}

```

emgGaussian_guess	<i>Guess function for initial exponentially modified gaussian parameters and bounds</i>
-------------------	---

Description

Guess function for initial exponentially modified gaussian parameters and bounds, at the moment only checks the x position

Usage

```
emgGaussian_guess(x, y)
```

Arguments

x (numeric) x values (e.g. retention time)
 y (numeric) y observed values (e.g. spectra intensity)

Value

A list of guessed starting parameters `list()`\$init_params, lower `list()`\$lower_bounds and upper bounds `list()`\$upper_bounds (`$gamma`, `$center`, `$sigma`, `$amplitude`)

emgGaussian_minpack.lm

Implementation of the Exponentially Modified Gaussian (EMG) peak shape for use with minpack.lm

Description

Implementation of the Exponentially Modified Gaussian (EMG) peak shape for use with minpack.lm

Usage

emgGaussian_minpack.lm(params, xx)

Arguments

params (list) exponential modified gaussian parameters (`params$gamma`, `params$center`, `params$sigma`, `params$amplitude`)
 xx (numeric) values at which to evaluate the exponentially modified gaussian

Value

value of the exponentially modified gaussian evaluated at xx

emgGaussian_minpack.lm_objectiveFun

Exponentially Modified Gaussian minpack.lm objective function

Description

Exponentially Modified Gaussian (EMG) minpack.lm objective function, calculates residuals using the EMG Peak Shape

Usage

emgGaussian_minpack.lm_objectiveFun(params, observed, xx)

Arguments

params	(list) exponentially modified gaussian parameters (params\$gamma, params\$center, params\$sigma, params\$amplitude)
observed	(numeric) observed y value at xx
xx	(numeric) value at which to evaluate the exponentially modified gaussian

Value

difference between observed and expected exponentially modified gaussian value evaluated at xx

extractSignalRawData *Extract signal in a multiple defined mz rt window from a raw data file*

Description

Extract all signal from multiple defined mz rt window from raw data and returns a data.frame. If no rt-mz window is provided, all signal in the raw data file are returned

Usage

```
extractSignalRawData(rawSpec, rt, mz, msLevel = 1L, verbose = TRUE)
```

Arguments

rawSpec	an <code>OnDiskMSnExp-class</code>
rt	(numeric(2) or two-column matrix) the lower and upper retention time range from which the data should be extracted. If a matrix is passed, each row corresponds to a different window. If not provided, the full retention time range will be extracted.
mz	(numeric(2) or two-column matrix) the lower and upper mass range from which the data should be extracted. If a matrix is passed, each row corresponds to a different window. If not provided, the full mass range will be extracted.
msLevel	(int) the MS level at which the data should be extracted (default to MS level 1)
verbose	(bool) If TRUE message progress and warnings

Details

```
## Examples cannot be computed as the function is not exported: ## Use a file from the faahKO
package and extract data from a region of ## interest library(faahKO) rawSpec <- MSnbase::readMSData(
system.file('cdf/KO/ko15.CDF',package='faahKO'), centroided=TRUE, mode='onDisk') dataPoints
<- extractSignalRawData(rawSpec, rt = c(3290., 3410.), mz = c(522.194778, 522.205222), ver-
bose=TRUE) # Reading data from 1 windows
```

```
dataPoints # [[1]] # rt mz int # 1 3290.115 522.2 1824 # 2 3291.680 522.2 1734 # 3 3293.245 522.2
1572 # 4 3294.809 522.2 1440 # 5 3299.504 522.2 1008 # 6 3301.069 522.2 871 # 7 3302.634
522.2 786 # 8 3304.199 522.2 802 # 9 3305.764 522.2 834 # 10 3307.329 522.2 839 # 11 3315.154
522.2 2187 # 12 3316.719 522.2 3534 # 13 3318.284 522.2 6338 # 14 3319.849 522.2 11718 # 15
3321.414 522.2 21744 # 16 3322.979 522.2 37872 # 17 3324.544 522.2 62424 # 18 3326.109 522.2
98408 # 19 3327.673 522.2 152896 # 20 3329.238 522.2 225984 # 21 3330.803 522.2 308672 # 22
3332.368 522.2 399360 # 23 3333.933 522.2 504000 # 24 3335.498 522.2 614656 # 25 3337.063
```

```
522.2 711872 # 26 3338.628 522.2 784704 # 27 3340.193 522.2 836608 # 28 3341.758 522.2
866304 # 29 3343.323 522.2 882304 # 30 3344.888 522.2 889280 # 31 3346.453 522.2 888256 # 32
3348.018 522.2 866816 # 33 3349.583 522.2 827392 # 34 3351.148 522.2 777728 # 35 3352.713
522.2 727040 # 36 3354.278 522.2 678464 # 37 3355.843 522.2 629120 # 38 3357.408 522.2
578048 # 39 3358.973 522.2 524288 # 40 3360.538 522.2 471040 # 41 3362.102 522.2 416320 #
42 3363.667 522.2 360064 # 43 3365.232 522.2 302400 # 44 3366.797 522.2 249152 # 45 3368.362
522.2 202560 # 46 3369.927 522.2 161024 # 47 3371.492 522.2 123520 # 48 3373.057 522.2 93160
# 49 3374.622 522.2 71856 # 50 3376.187 522.2 58392 # 51 3377.752 522.2 51072 # 52 3379.317
522.2 48376 # 53 3380.882 522.2 49168 # 54 3382.447 522.2 53120 # 55 3384.012 522.2 62488 #
56 3385.577 522.2 78680 # 57 3387.142 522.2 102840 # 58 3388.707 522.2 134656 # 59 3390.272
522.2 173440 # 60 3391.837 522.2 217088 # 61 3393.402 522.2 268864 # 62 3394.966 522.2
330496 # 63 3396.531 522.2 395776 # 64 3398.096 522.2 453376 # 65 3399.661 522.2 499072 #
66 3401.226 522.2 537024 # 67 3402.791 522.2 570304 # 68 3404.356 522.2 592512 # 69 3405.921
522.2 598912 # 70 3407.486 522.2 595008 # 71 3409.051 522.2 588416
```

Value

a list (one entry per window) of data.frame with signal as row and retention time ('rt'), mass ('mz') and intensity ('int') as columns.

```
filename,peakPantheRAnnotation-method
      filename accessor by splitting filepath
```

Description

filename accessor by splitting filepath

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
filename(object)
```

Arguments

object peakPantheRAnnotation

Value

(str) filename

Examples

```
if(requireNamespace('faahK0')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahK0)
  spectraPaths <- c(system.file('cdf/K0/ko15.CDF', package = 'faahK0'),
                    system.file('cdf/K0/ko16.CDF', package = 'faahK0'),
                    system.file('cdf/K0/ko18.CDF', package = 'faahK0'))
```

```

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

filename(annotation)
# [1] 'ko15' 'ko16' 'ko18'
}

```

```

filepath,peakPantheRAnnotation-method
  filepath accessor

```

Description

filepath accessor

Usage

```

## S4 method for signature 'peakPantheRAnnotation'
filepath(object)

```

Arguments

object peakPantheRAnnotation

Value

(str) A character vector of file paths, of length number of spectra files

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',

```

```

      'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                        522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                        496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
                                   FUN.VALUE=numeric(2))

annotation <- peakPantherAnnotation(spectraPaths=spectraPaths,
                                   targetFeatTable=targetFeatTable)

filepath(annotation)
# [1] 'C:/R/R-3.6.0/library/faahKO/cdf/KO/ko15.CDF'
# [2] 'C:/R/R-3.6.0/library/faahKO/cdf/KO/ko16.CDF'
# [3] 'C:/R/R-3.6.0/library/faahKO/cdf/KO/ko18.CDF'
}

```

findTargetFeatures *Find and integrate target features in each ROI*

Description

For each ROI, fit a curve and integrate the largest feature in the box. Each entry in ROIsDataPoints must match the corresponding row in ROI. The curve shape to employ for fitting can be changed with curveModel while fitting parameters can be changed with params (list with one param per ROI window). rtMin and rtMax are established at 0.5 outward (the window is the ROI width); if after 8 iterations rtMin or rtMax is not found, NA is returned and the peak fit rejected. peakArea is calculated from rtMin to rtMax. peakAreaRaw is calculated from rtMin to rtMax but using the raw data points instead of the modelled line-shape. mz is the weighted (by intensity) average mz of datapoints falling into the rtMin to rtMax range, mzMin and mzMax are the minimum and maximum mass in these range. If rtMin or rtMax falls outside of ROI (extracted scans), mzMin or mzMax are returned as the input ROI limits and mz is an approximation on the datapoints available (if no scan of the ROI fall between rtMin/rtMax, mz would be NA, the peak is rejected). If any of the two following ratio are superior to maxApexResidualRatio, the fit is rejected: 1) ratio of fit residuals at the apex (predicted apex fit intensity vs measured apex intensity: fit overshoots the apex), 2) ratio of predicted apex fit intensity vs maximum measured peak intensity (fit misses the real apex in the peak).

Usage

```

findTargetFeatures(
  ROIsDataPoints,
  ROI,
  curveModel = "skewedGaussian",
  params = "guess",
  sampling = 250,
  maxApexResidualRatio = 0.2,
  verbose = FALSE,
  ...
)

```

Arguments

ROIsDataPoints	(list) A list (one entry per ROI window) of data.frame with signal as row and retention time ('rt'), mass ('mz') and intensity ('int') as columns. Must match each row of ROI.
ROI	(data.frame) A data.frame of compounds to target as rows. Columns: rtMin (float in seconds), rtMax (float in seconds), mzMin (float), mzMax (float)
curveModel	(str) Name of the curve model to fit (currently skewedGaussian and emgGaussian)
params	(list or str) Either 'guess' for automated parametrisation or list (one per ROI windows) of 'guess' or list of curve fit parameters
sampling	(int) Number of points to employ when subsampling the fittedCurve (rt, rtMin, rtMax, integral calculation)
maxApexResidualRatio	(float) Ratio of maximum allowed fit residual at the peak apex, compared to the fit max intensity. (e.g. 0.2 for a maximum residual of 20% of apex intensity)
verbose	(bool) If TRUE message the time taken and number of features found
...	Passes arguments to fitCurve to alter peak fitting (params)

Details

```
## Examples cannot be computed as the function is not exported: ## Load data library(faahKO) li-
brary(MSNbase) netcdfFilePath <- system.file('cdf/KO/ko15.CDF', package = 'faahKO') raw_data
<- MSNbase::readMSData(netcdfFilePath,centroided=TRUE,mode='onDisk')
```

```
## targetFeatTable targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(), c('cpdID', 'cpdName', 'rtMin',
'mz', 'mzMax'))), stringsAsFactors=FALSE) targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888,
3390., 522.194778, 522.2, 522.205222) targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577,
3440., 496.195038, 496.2, 496.204962) targetFeatTable[,3:8] <- vapply(targetFeatTable[,3:8], as.numeric,
FUN.VALUE=numeric(2))
```

```
ROIsPt <- extractSignalRawData(raw_data, rt=targetFeatTable[,c('rtMin', 'rtMax')], mz=targetFeatTable[,c('mzMin', 'mzMax')],
verbose=TRUE) # Reading data from 2 windows
```

```
foundPeaks <- findTargetFeatures(ROIsPt, targetFeatTable, verbose=TRUE) # Warning: rtMin/rtMax
outside of ROI; datapoints cannot be used for # mzMin/mzMax calculation, # approximate mz and
returning ROI$mzMin and ROI$mzMax for ROI #1 # Found 2/2 features in 0.07 secs
```

```
foundPeaks # $peakTable # found rtMin rt rtMax mzMin mz mzMax peakArea # 1 TRUE 3309.759
3346.828 3385.410 522.1948 522.2 522.2052 26133727 # 2 TRUE 3345.377 3386.529 3428.279
496.2000 496.2 496.2000 35472141 # peakAreaRaw maxIntMeasured maxIntPredicted # 1 26071378
889280 901015.8 # 2 36498367 1128960 1113576.7 # # $curveFit # $curveFit[[1]] # $amplitude #
[1] 162404.8 # # $center # [1] 3341.888 # # $sigma # [1] 0.07878613 # # $gamma # [1] 0.00183361
# # $fitStatus # [1] 2 # # $curveModel # [1] 'skewedGaussian' # # attr('class') # [1] 'peakPantheR_curveFit'
# # $curveFit[[2]] # $amplitude # [1] 199249.1 # # $center # [1] 3382.577 # #
$sigma # [1] 0.07490442 # # $gamma # [1] 0.00114719 # # $fitStatus # [1] 2 # # $curveModel #
[1] 'skewedGaussian' # # attr('class') # [1] 'peakPantheR_curveFit'
```

Value

A list: list()\$peakTable (*data.frame*) with targeted features as rows and peak measures as columns (see Details), list()\$curveFit (*list*) a list of peakPantheR_curveFit or NA for each ROI.

Details:: The returned data.frame is structured as follow:

found	was the peak found
rt	retention time of peak apex (sec)
rtMin	leading edge of peak retention time (sec) determined at 0.5% of apex intensity
rtMax	trailing edge of peak retention time (sec) determined at 0.5% of apex intensity
mz	weighted (by intensity) mean of peak m/z across scans
mzMin	m/z peak minimum (between rtMin, rtMax)
mzMax	m/z peak maximum (between rtMin, rtMax)
peakArea	integrated peak area
peakAreaRaw	integrated peak area from raw data points
maxIntMeasured	maximum peak intensity in raw data
maxIntPredicted	maximum peak intensity based on curve fit (at apex)

FIR,peakPantheRAnnotation-method

FIR accessor returns targetFeatTable with cpdID, cpdName added

Description

FIR accessor returns targetFeatTable with cpdID, cpdName added

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
FIR(object)
```

Arguments

object peakPantheRAnnotation

Value

(data.frame) target feature table with compounds as row and FIR parameters as columns

Examples

```
if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
'mzMax'))), stringsAsFactors=FALSE)
```



```

targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                        522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                        496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
                                  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
                                   targetFeatTable=targetFeatTable)

## default values without annotation
FIR(annotation)
#   rtMin rtMax mzMin mzMax cpdID cpdName
# 1   NA   NA   NA   NA   ID-1   Cpd 1
# 2   NA   NA   NA   NA   ID-2   Cpd 2
}

```

fitCurve

Curve fitting using minpack.lm

Description

Fit different curve models using minpack. Fitting parameters can be passed or guessed.

Usage

```
fitCurve(x, y, curveModel = "skewedGaussian", params = "guess")
```

Arguments

x	(numeric) x values (e.g. retention time)
y	(numeric) y observed values (e.g. spectra intensity)
curveModel	(str) name of the curve model to fit (currently skewedGaussian and emgGaussian)
params	(list or str) either 'guess' for automated parametrisation or list of initial parameters (\$init_params), lower parameter bounds (\$lower_bounds) and upper parameter bounds (\$upper_bounds)

Details

```

## Examples cannot be computed as the function is not exported: ## x is retention time, y corresponding intensity
input_x <- c(3362.102, 3363.667, 3365.232, 3366.797, 3368.362, 3369.927, 3371.492, 3373.057, 3374.622, 3376.187, 3377.752, 3379.317, 3380.882, 3382.447, 3384.012, 3385.577, 3387.142, 3388.707, 3390.272, 3391.837, 3393.402, 3394.966, 3396.531, 3398.096, 3399.661, 3401.226, 3402.791, 3404.356, 3405.921, 3407.486, 3409.051)
input_y <- c(51048, 81568, 138288, 233920, 376448, 557288, 753216, 938048, 1091840, 1196992, 1261056, 1308992, 1362752, 1406592, 1431360, 1432896, 1407808, 1345344, 1268480, 1198592, 1126848, 1036544, 937600, 849792, 771456, 692416, 614528, 546088, 492752, 446464, 400632)

## Fit fitted_curve <- fitCurve(input_x, input_y, curveModel='skewedGaussian', params='guess')
## Returns the optimal fitting parameters fitted_curve ## $amplitude # [1] 275371.1 ## $center # [1] 3382.577 ## $sigma # [1] 0.07904697 ## $gamma # [1] 0.001147647 ## $fitStatus # [1] 2 ## $curveModel # [1] 'skewedGaussian' ## attr(,"class") # [1] 'peakPantheR_curveFit'

```

Value

A 'peakPantheR_curveFit': a list of fitted curve parameters, fitStatus from nls.lm\$info and curve shape name curveModel. fitStatus=0 unsuccessful completion: improper input parameters, fitStatus=1 successful completion: first convergence test is successful, fitStatus=2 successful completion: second convergence test is successful, fitStatus=3 successful completion: both convergence test are successful, fitStatus=4 questionable completion: third convergence test is successful but should be carefully examined (maximizers and saddle points might satisfy), fitStatus=5 unsuccessful completion: excessive number of function evaluations/iterations

gaussian_cerf	<i>Gaussian Error function</i>
---------------	--------------------------------

Description

Implementation of the gaussian error function

Usage

```
gaussian_cerf(x)
```

Arguments

x (numeric) value at which to evaluate the gaussian error function

Value

Value of the gaussian error function evaluated at x

gaussian_erf	<i>Gaussian Error function</i>
--------------	--------------------------------

Description

Implementation of the gaussian error function

Usage

```
gaussian_erf(x)
```

Arguments

x (numeric) value at which to evaluate the gaussian error function

Value

Value of the gaussian error function evaluated at x

```
generateIonChromatogram
```

Generate ion chromatogram from raw data points

Description

On the input raw data, aggregate intensities across the m/z range at each retention time to generate an ion chromatogram: sum for EIC/TIC, max, min or mean. The number of data points returned correspond to the number of unique scans/retention time measurements in the input data

Usage

```
generateIonChromatogram(ROIDataPoint, aggregationFunction = "sum")
```

Arguments

ROIDataPoint (data.frame) retention time 'rt', mass 'mz' and intensity 'int' (as column) of each raw data points (as row) to use for the ion chromatogram

aggregationFunction (str) Function to use in order to aggregate intensities across m/z in each scan. One of sum, max, min, mean

Details

```
## Examples cannot be computed as the function is not exported: ## Input data points in_rt <-
c(3362.102, 3362.102, 3363.667, 3363.667, 3365.232, 3365.232, 3366.797, 3366.797, 3368.362,
3368.362) in_mz <- c(496.2, 497.2, 496.2, 497.2, 496.2, 497.2, 496.2, 497.2, 496.2, 497.2) in_int <-
c(39616, 11432, 63344, 18224, 107352, 30936, 182144, 51776, 295232, 81216) input_ROIDataPoints
<- data.frame(rt=in_rt, mz=in_mz, int=in_int)
```

```
## Aggregate m/z to generate EIC EIC <- generateIonChromatogram(input_ROIDataPoints,aggregationFunction='sum')
EIC # rt int # 1 3362.102 51048 # 2 3363.667 81568 # 3 3365.232 138288 # 4 3366.797 233920 #
5 3368.362 376448
```

Value

A data.frame of retention time 'rt' and aggregated intensities 'int'

```
getAcquisitionDateMzML
```

Parse acquisition date from a mzML file

Description

Extract acquisition date ("startTimeStamp") from a mzML file. In case of failure (or the file is not a mzML) returns NULL

Usage

```
getAcquisitionDateMzML(mzMLPath, verbose = TRUE)
```

Arguments

mzMLPath (str) path to mzML raw data file
 verbose (bool) if TRUE message progress

Value

POSIXct or NA

getTargetFeatureStatistic

Calculate chromatographic peak properties

Description

Calculate the ppm error, retention time deviation, tailing factor and asymmetry factor for each measured feature.

Usage

```
getTargetFeatureStatistic(
  fittedCurves,
  targetFeatTable,
  foundPeakTable,
  verbose = FALSE
)
```

Arguments

fittedCurves (list) A list (one entry per ROI window) of peakPantheR_curveFit or NA
 targetFeatTable a [data.frame](#) of compounds to target as rows. Columns: cpdID (str), cpdName (str), rtMin (float in seconds), rt (float in seconds, or NA), rtMax (float in seconds), mzMin (float), mz (float or NA), mzMax (float).
 foundPeakTable a [data.frame](#) as generated by [findTargetFeatures](#), with features as rows and peak properties as columns. The following columns are mandatory: mzMin, mz, mzMax, rtMin, rt, rtMax
 verbose (bool) if TRUE message when NA scans are removed

Details

```
## Examples cannot be computed as the function is not exported: # fittedCurve cFit1 <- list(amplitude=162404.80579182,
center=3341.888, sigma=0.078786133031045896, gamma=0.0018336101984172684, fitStatus=2,
curveModel='skewedGaussian') class(cFit1) <- 'peakPantheR_curveFit' cFit2 <- list(amplitude=199249.10572753669,
center=3382.577, sigma=0.074904415304607966, gamma=0.0011471899372353885, fitStatus=2,
curveModel='skewedGaussian') class(cFit2) <- 'peakPantheR_curveFit' input_fitCurves <- list(cFit1,
cFit2)

# ROI input_ROI <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(), c('cpdID', 'cpdName', 'rt-
Min', 'rt', 'rtMax', 'mzMin', 'mz', 'mzMax'))), stringsAsFactors=FALSE) input_ROI[1,] <- c('ID-
1', 'testCpd 1', 3310., 3344.88, 3390., 522.19, 522.2, 522.21) input_ROI[2,] <- c('ID-2', 'testCpd
```

```

2', 3280., 3385.58, 3440., 496.19, 496.2, 496.21) input_ROI[,3:8] <- vapply(input_ROI[,3:8], as.numeric,
FUN.VALUE=numeric(2))

# foundPeakTable input_foundPeakTable <- data.frame(matrix(vector(), 2, 10, dimnames=list(c(),
c('found', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz', 'mzMax', 'peakArea', 'maxIntMeasured', 'maxIntPredicted'))),
stringsAsFactors=FALSE) input_foundPeakTable[1,] <- c(TRUE, 3309.758, 3346.827, 3385.410,
522.19, 522.2, 522.21, 26133726, 889280, 901015) input_foundPeakTable[2,] <- c(TRUE, 3345.376,
3386.529, 3428.279, 496.19, 496.2, 496.21, 35472141, 1128960, 1113576) input_foundPeakTable[,1]
<- vapply(input_foundPeakTable[,c(1)], as.logical, FUN.VALUE=logical(1))

# Run peak statistics peakStatistics <- getTargetFeatureStatistic(input_fitCurves, input_ROI, in-
put_foundPeakTable) peakStatistics # found rtMin rtMax mzMin mz mzMax peakArea # 1 TRUE
3309.758 3346.827 3385.410 522.19 522.2 522.21 26133726 # 2 TRUE 3345.376 3386.529 3428.279
496.19 496.2 496.21 35472141 # maxIntMeasured maxIntPredicted ppm_error rt_dev_sec tailing-
Factor # 1 889280 901015 0 1.947 1.015385 # 2 1128960 1113576 0 0.949 1.005372 # asymmetry-
Factor # 1 1.026886 # 2 1.009304

```

Value

A data.frame with measured compounds as rows and measurements and properties as columns (see Details).

Details:: The returned data.frame is structured as follow:

found	was the peak found
rt	retention time of peak apex (sec)
rtMin	leading edge of peak retention time (sec) determined at 0.5% of apex intensity
rtMax	trailing edge of peak retention time (sec) determined at 0.5% of apex intensity
mz	weighted (by intensity) mean of peak m/z across scans
mzMin	m/z peak minimum (between rtMin, rtMax)
mzMax	m/z peak maximum (between rtMin, rtMax)
peakArea	integrated peak area
peakAreaRaw	integrated peak area from raw data points
maxIntMeasured	maximum peak intensity in raw data
maxIntPredicted	maximum peak intensity based on curve fit
ppm_error	difference in ppm between the expected and measured m/z
rt_dev_sec	difference in seconds between the expected and measured rt
tailingFactor	the tailing factor is a measure of peak tailing. It is defined as the distance from the front slope of the p
asymmetryFactor	the asymmetry factor is a measure of peak tailing. It is defined as the distance from the center line of t

```
initialise_annotation_from_files_UI_helper
```

UI data import helper - initialise new annotation from files

Description

Fully initialise a peakPantherAnnotation using the target files path, CSV parameter path and metadata.

Usage

```
initialise_annotation_from_files_UI_helper(
  CSVParamPath,
  spectraPaths,
  cpdMetadataPath = NULL,
  spectraMetadata = NULL,
  verbose = TRUE
)
```

Arguments

CSVParamPath (str) Path to a CSV file of fit parameters

spectraPaths (str) character vector of spectra file paths, to set samples to process

cpdMetadataPath
NULL or path to a csv of compound metadata, with compounds as row and metadata as columns

spectraMetadata
NULL or DataFrame of sample metadata, with samples as row and metadata as columns

verbose (bool) If TRUE message progress

Value

(peakPantheRAnnotation) Object initialised with ROI, uROI and FIR read from the CSV file

Examples

```
## Input data
input_CSV <- data.frame(matrix(nrow=2,ncol=21,dimnames=list(c(),
  c('cpdID', 'cpdName',
    'X','ROI_rt', 'ROI_mz', 'ROI_rtMin', 'ROI_rtMax','ROI_mzMin','ROI_mzMax',
    'X','uROI_rtMin', 'uROI_rtMax', 'uROI_mzMin', 'uROI_mzMax', 'uROI_rt',
    'uROI_mz', 'X', 'FIR_rtMin', 'FIR_rtMax', 'FIR_mzMin', 'FIR_mzMax'))))
input_CSV[1,] <- c('ID-1', 'Cpd 1', '|', 1., 2., 3., 4., 5., 6., '|',
  7., 8., 9., 10., 11., 12., '|', 13., 14., 15., 16.)
input_CSV[2,] <- c('ID-2', 'Cpd 2', '|', 17., 18., 19., 20., 21., 22., '|',
  23., 24., 25., 26., 27., 28., '|', 29., 30., 31., 32.)
input_CSV[,-c(1,2,3,10,17)] <- vapply(input_CSV[,-c(1,2,3,10,17)],
  as.numeric, FUN.VALUE=numeric(2))

input_spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# temporary file location
savePath1 <- tempfile(pattern='file', tmpdir=tempdir(), fileext='.csv')
# save csv
utils::write.csv(input_CSV, file=savePath1, row.names=FALSE)

# Load parameters from CSV
loadedAnnotation <- initialise_annotation_from_files_UI_helper(savePath1,
  input_spectraPaths,
  verbose=TRUE)

# An object of class peakPantheRAnnotation
# 2 compounds in 3 samples.
# updated ROI exist (uROI)
```

```
# does not use updated ROI (uROI)
# does not use fallback integration regions (FIR)
# is not annotated
```

integrateFIR *Integrate fallback integration regions*

Description

Integrate region defined in FIR if a feature is not found

Usage

```
integrateFIR(rawSpec, FIR, foundPeakTable, verbose = TRUE)
```

Arguments

rawSpec	an OnDiskMSnExp-class
FIR	(data.frame) Fallback Integration Regions (FIR) to integrate when a feature is not found. Compounds as row are identical to the targeted features, columns are rtMin (float in seconds), rtMax (float in seconds), mzMin (float), mzMax (float)
foundPeakTable	a data.frame as generated by findTargetFeatures , with features as rows and peak properties as columns. The following columns are mandatory: found, is_filled, mz, mzMin, mzMax, rt, rtMin, rtMax, peakArea, peakAreaRaw, maxIntMeasured, maxIntPredicted.
verbose	(bool) if TRUE message progress

Value

an updated foundPeakTable with FIR integration values

is.peakPantheR_curveFit
Check if object is of class peakPantheR_curveFit

Description

Check if object is of class peakPantheR_curveFit

Usage

```
is.peakPantheR_curveFit(x)
```

Arguments

x	object to test
---	----------------

Value

(bool) TRUE or FALSE

isAnnotated,peakPantheRAnnotation-method
isAnnotated accessor

Description

isAnnotated accessor

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
isAnnotated(object)
```

Arguments

object peakPantheRAnnotation

Value

(bool) flag if the annotation has taken place

Examples

```
if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
targetFeatTable=targetFeatTable)

isAnnotated(annotation)
# [1] FALSE
}
```

`load_annotation_from_file_UI_helper`*UI data import helper - check loaded annotation*

Description

Load a .RData file (check it exists) and that a peakPantheRAnnotation named "annotationObject" is present. Returns the annotation if everything is valid

Usage

```
load_annotation_from_file_UI_helper(annotationPath)
```

Arguments

`annotationPath` (str) Path to a RData file containing a peakPantheRAnnotation names 'annotationObject'

Value

(peakPantheRAnnotation) Object loaded from file

Examples

```
## Initialise a peakPantheRAnnotation object with 3 samples and 2 compounds

## Inputs
# spectraPaths
spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

annotationObject <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# save annotation to disk
annotPath <- tempfile(pattern="file", tmpdir=tempdir(), fileext='.RData')
save(annotationObject, file=annotPath, compress=TRUE)

# Load annotation
load_annotation_from_file_UI_helper(annotationPath = annotPath)
# An object of class peakPantheRAnnotation
# 2 compounds in 3 samples.
# updated ROI do not exist (uROI)
# does not use updated ROI (uROI)
```

```
# does not use fallback integration regions (FIR)
# is not annotated
```

```
nbCompounds, peakPantheRAnnotation-method
      nbCompounds accessor established on cpdID
```

Description

nbCompounds accessor established on cpdID

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
nbCompounds(object)
```

Arguments

object peakPantheRAnnotation

Value

(int) number of samples

Examples

```
if(requireNamespace('faahKO')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahKO)
  spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                    system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

  # targetFeatTable
  targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
    c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
  targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
    522.2, 522.205222)
  targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
    496.2, 496.204962)
  targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
    FUN.VALUE=numeric(2))

  annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
    targetFeatTable=targetFeatTable)

  nbCompounds(annotation)
  # [1] 2
}
```

nbSamples, peakPantheRAnnotation-method
nbSamples accessor established on filepath

Description

nbSamples accessor established on filepath

Usage

```
## S4 method for signature 'peakPantheRAnnotation'  
nbSamples(object)
```

Arguments

object peakPantheRAnnotation

Value

(int) number of samples

Examples

```
if(requireNamespace('faahKO')){  
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted  
  ## compounds  
  
  # Paths to spectra files  
  library(faahKO)  
  spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),  
                    system.file('cdf/KO/ko16.CDF', package = 'faahKO'),  
                    system.file('cdf/KO/ko18.CDF', package = 'faahKO'))  
  
  # targetFeatTable  
  targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),  
    c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',  
    'mzMax'))), stringsAsFactors=FALSE)  
  targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,  
    522.2, 522.205222)  
  targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,  
    496.2, 496.204962)  
  targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,  
    FUN.VALUE=numeric(2))  
  
  annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,  
    targetFeatTable=targetFeatTable)  
  
  nbSamples(annotation)  
  # [1] 3  
}
```

```
outputAnnotationDiagnostic,peakPantheRAnnotation-method
```

Save to disk the annotation parameters as CSV and a diagnostic plot per fitted compound

Description

Save to disk the annotation parameters as CSV (as generated by `outputAnnotationParamsCSV()`) and a diagnostic plot per fitted compound (as generated by `annotationDiagnosticMultiplot()`) if `savePlots` is TRUE

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
outputAnnotationDiagnostic(
  object,
  saveFolder,
  savePlots = TRUE,
  sampleColour = NULL,
  verbose = TRUE,
  ncores = 0,
  svgPlot = FALSE,
  ...
)
```

Arguments

<code>object</code>	(<code>peakPantheRAnnotation</code>) Annotated <code>peakPantheRAnnotation</code> object
<code>saveFolder</code>	(<code>str</code>) Path of folder where <code>annotationParameters_summary.csv</code> and plots will be saved
<code>savePlots</code>	(<code>bool</code>) If TRUE save a diagnostic plot for each compound
<code>sampleColour</code>	(<code>str</code>) NULL or vector colour for each sample
<code>verbose</code>	(<code>bool</code>) If TRUE message progress
<code>ncores</code>	(<code>int</code>) Number of cores to use to save plots in parallel
<code>svgPlot</code>	(<code>bool</code>) If TRUE save plots as 'svg', otherwise as 'png'
<code>...</code>	Additional parameters for plotting i.e. <code>sampling</code> for the number of points to employ when plotting <code>fittedCurve</code>

Value

None

Examples

```
if(requireNamespace('faahKO')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahKO)
```

```
spectraPaths <- c(system.file('cdf/K0/ko15.CDF', package = 'faahK0'),
                  system.file('cdf/K0/ko16.CDF', package = 'faahK0'),
                  system.file('cdf/K0/ko18.CDF', package = 'faahK0'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# Calculate annotation
annotation <- peakPantheR_parallelAnnotation(emptyAnnotation, ncores=0,
  getAcqTime=FALSE, verbose=FALSE)$annotation

# temporary location
savePath1 <- tempdir()
outputAnnotationDiagnostic(annotation, saveFolder=savePath1, savePlots=FALSE,
  verbose=TRUE)
}
```

outputAnnotationFeatureMetadata_UI_helper

UI export helper - feature metadata

Description

Return a table with features as rows and all feature metadata as columns

Usage

```
outputAnnotationFeatureMetadata_UI_helper(annot)
```

Arguments

annot (peakPantheRAnnotation) Annotation object

Value

(data.frame) Features metadata

Examples

```
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
```

```

spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# featureMetadata of an empty annotation
outputAnnotationFeatureMetadata_UI_helper(emptyAnnotation)
# data frame with 0 columns and 2 rows

```

```

outputAnnotationParamsCSV,peakPantheRAnnotation-method
  Save annotation parameters as CSV

```

Description

Save annotation parameters (ROI, uROI and FIR) to disk as a CSV file for editing

Usage

```

## S4 method for signature 'peakPantheRAnnotation'
outputAnnotationParamsCSV(
  object,
  saveFolder,
  verbose,
  noSave
)

```

Arguments

object	(peakPantheRAnnotation) Annotated peakPantheRAnnotation object
saveFolder	(str) Path of folder where annotationParameters_summary.csv will be saved
verbose	(bool) If TRUE message progress
noSave	(bool) If TRUE the resulting table will be returned without saving to disk

Value

None

Examples

```
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# temporary file
savePath <- tempdir()

# statistics of an empty annotation
outputAnnotationParamsCSV(emptyAnnotation, saveFolder=savePath, verbose=TRUE)
```

```
outputAnnotationResult,peakPantheRAnnotation-method
```

Save to disk all annotation results as csv files

Description

Save to disk all annotation results as annotationName_... .csv files: compound metadata (cpdMetadata, cpdID, cpdName) and spectra metadata (spectraMetadata, acquisitionTime, TIC), summary of fit (ratio of peaks found: ratio_peaks_found, ratio of peaks filled: ratio_peaks_filled, mean ppm_error: ppm_error, mean rt_dev_sec: rt_dev_sec), and a file for each column of peakTables (with samples as rows and compounds as columns)

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
outputAnnotationResult(
  object,
  saveFolder,
  annotationName = "annotationResult",
  verbose = TRUE
)
```

Arguments

object (peakPantheRAnnotation) Annotated peakPantheRAnnotation object

saveFolder (str) Path of folder where the annotation result csv will be saved
 annotationName (str) name of annotation to use in the saved csv
 verbose (bool) If TRUE message progress

Value

None

Examples

```
if(requireNamespace('faahKO')){
## Initialise a peakPantherAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantherAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# Calculate annotation
annotation <- peakPanther_parallelAnnotation(emptyAnnotation, ncores=0,
  getAcqTime=FALSE, verbose=FALSE)$annotation

# temporary location
savePath1 <- tempdir()
outputAnnotationResult(annotation, saveFolder=savePath1,
  annotationName='testProject', verbose=TRUE)
}
```

outputAnnotationSpectraMetadata_UI_helper

UI export helper - spectra path and metadata

Description

Return a table with spectra as rows and filepath and all spectra metadata columns

Usage

```
outputAnnotationSpectraMetadata_UI_helper(annot)
```

Arguments

```
annot          (peakPantheRAnnotation) Annotation object
```

Value

```
(data.frame) Spectra paths and metadata
```

Examples

```
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# spectraMetada of an empty annotation
outputAnnotationSpectraMetadata_UI_helper(emptyAnnotation)
#      filepath
# 1 ./path/file1
# 2 ./path/file2
# 3 ./path/file3
```

```
peakFit,peakPantheRAnnotation-method
      peakFit accessor
```

Description

```
peakFit accessor
```

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
peakFit(object)
```

Arguments

object peakPantheRAnnotation

Value

A list of length number of spectra files. Each list element is a *curveFit* list of peakPantheR_curveFit or NA for each ROI

Examples

```
if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

## default values without annotation
peakFit(annotation)
# [[1]]
# NULL
# [[2]]
# NULL
# [[3]]
# NULL
}
```

peakPantheR

peakPantheR: A package for Peak Picking and ANnotation of High resolution Experiments

Description

peakPantheR detects, integrates and reports pre-defined features in mass spectrometry data files. It enables the real time annotation of multiple compounds in a single file, or the parallel annotation of multiple compounds in multiple files.

Details

The main functions of **peakPantheR** are [peakPantheR_singleFileSearch](#) for realtime annotation, and [peakPantheR_parallelAnnotation](#) for parallel annotation. The `peakPantheRAnnotation` object stores parallel annotation results, while reporting functions help assess the quality of annotation and update fitting parameters. Refer to the vignettes for graphical user interface and command line tutorials.

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See Also

Useful links:

- <https://github.com/phenomecentre/peakPantheR>
- Report bugs at <https://github.com/phenomecentre/peakPantheR/issues/new>

peakPantheRAnnotation *An S4 class to represent peakPantheR annotation results*

Description

The `peakPantheRAnnotation` class is designed to run and store `peakPantheR` parallel annotation results. Instances of the class are created with the `peakPantheRAnnotation` constructor function, which initialises an object of proper dimension with `spectraPaths` (set samples to process) and `targetFeatTable` (set compounds to target). `spectraPaths` is a character vector of spectra file paths. `targetFeatTable` is a [data.frame](#) of compounds to target as rows and parameters as columns: `cpdID` (int), `cpdName` (str), `rtMin` (float in seconds), `rt` (float in seconds, or *NA*), `rtMax` (float in seconds), `mzMin` (float), `mz` (float or *NA*), `mzMax` (float).

`peakPantheRAnnotation()`: create an instance of the `peakPantherAnnotation` class.

Usage

```
peakPantheRAnnotation(spectraPaths = NULL, targetFeatTable = NULL,
  cpdID = character(), cpdName = character(), ROI = data.frame(rtMin =
  numeric(), rt = numeric(), rtMax = numeric(), mzMin = numeric(), mz =
  numeric(), mzMax = numeric(), stringsAsFactors = FALSE),
  FIR = data.frame(rtMin = numeric(), rtMax = numeric(), mzMin =
  numeric(), mzMax = numeric(), stringsAsFactors = FALSE),
  uROI = data.frame(rtMin = numeric(), rt = numeric(), rtMax = numeric(),
  mzMin = numeric(), mz = numeric(), mzMax = numeric(), stringsAsFactors =
  FALSE), filepath = character(), cpdMetadata = data.frame(),
```

```
spectraMetadata = data.frame(), acquisitionTime = character(),
uROIExist = FALSE, useUROI = FALSE, useFIR = FALSE,
TIC = numeric(), peakTables = list(), dataPoints = list(),
peakFit = list(), isAnnotated = FALSE)
```

```
peakPantheRAnnotation(spectraPaths = NULL, targetFeatTable = NULL,
  cpdID = character(), cpdName = character(), ROI = data.frame(rtMin
= numeric(), rt = numeric(), rtMax = numeric(), mzMin = numeric(), mz =
numeric(), mzMax = numeric(), stringsAsFactors = FALSE),
  FIR = data.frame(rtMin = numeric(), rtMax = numeric(), mzMin =
numeric(), mzMax = numeric(), stringsAsFactors = FALSE),
  uROI = data.frame(rtMin = numeric(), rt = numeric(), rtMax = numeric(),
mzMin = numeric(), mz = numeric(), mzMax = numeric(), stringsAsFactors =
FALSE), filepath = character(), cpdMetadata = data.frame(),
  spectraMetadata = data.frame(), acquisitionTime = character(),
  uROIExist = FALSE, useUROI = FALSE, useFIR = FALSE,
  TIC = numeric(), peakTables = list(), dataPoints = list(),
  peakFit = list(), isAnnotated = FALSE)
```

Arguments

spectraPaths	NULL or a character vector of spectra file paths, to set samples to process
targetFeatTable	NULL or a data.frame of compounds to target as rows and parameters as columns: cpdID (str), cpdName (str), rtMin (float in seconds), rt (float in seconds, or NA), rtMax (float in seconds), mzMin (float), mz (float or NA), mzMax (float). Set compounds to target.
cpdID	A character vector of compound IDs, of length number of compounds
cpdName	A character vector of compound names, of length number of compounds
ROI	A data.frame of Regions Of Interest (ROI) with compounds as row and ROI parameters as columns: rtMin (float in seconds), rt (float in seconds, or NA), rtMax (float in seconds), mzMin (float), mz (float or NA), mzMax (float).
FIR	A data.frame of Fallback Integration Regions (FIR) with compounds as row and FIR parameters as columns: rtMin (float in seconds), rtMax (float in seconds), mzMin (float), mzMax (float).
uROI	A data.frame of updated Regions Of Interest (uROI) with compounds as row and uROI parameters as columns: rtMin (float in seconds), rt (float in seconds, or NA), rtMax (float in seconds), mzMin (float), mz (float or NA), mzMax (float).
filepath	A character vector of file paths, of length number of spectra files
cpdMetadata	A data.frame of compound metadata, with compounds as row and metadata as columns
spectraMetadata	A data.frame of sample metadata, with samples as row and metadata as columns
acquisitionTime	A character vector of acquisition date-time (converted from POSIXct) or NA
uROIExist	A logical stating if uROI have been set
useUROI	A logical stating if uROI are to be used
useFIR	A logical stating if FIR are to be used
TIC	A numeric vector of TIC or NA, of length number of spectra files

peakTables	A list of peakTable data.frame, of length number of spectra files. Each peakTable data.frame has compounds as rows and peak annotation results as columns.
dataPoints	A list of length number of spectra files. Each list element is a <i>ROIsDataPoint</i> list of data.frame of raw data points for each ROI/uROI (retention time 'rt', mass 'mz' and intensity 'int' (as column) of each raw data points (as row))
peakFit	A list of length number of spectra files. Each list element is a <i>curveFit</i> list of peakPantheR_curveFit or NA for each ROI
isAnnotated	A logical stating in the annotation took place

Details

The `validObject` method ensures the conformity of an object to the `peakPantheRAnnotation-class`. The number of compounds is based on `cpdID()` length, and the number of samples is based on `filepath()` length. Slot type is not checked as `setClass` enforces it. `peakTables` and `EICs` type are checked on the first list element. `annotationTable(object, column)` where *column* is a column from *peakTable*, returns a data.frame of values with the samples as rows, ROI as columns.

Value

(peakPantheRAnnotation)

Slots

cpdID	A character vector of compound IDs, of length number of compounds
cpdName	A character vector of compound names, of length number of compounds
ROI	A data.frame of Regions Of Interest (ROI) with compounds as row and ROI parameters as columns: <code>rtMin</code> (float in seconds), <code>rt</code> (float in seconds, or <i>NA</i>), <code>rtMax</code> (float in seconds), <code>mzMin</code> (float), <code>mz</code> (float or <i>NA</i>), <code>mzMax</code> (float).
FIR	A data.frame of Fallback Integration Regions (FIR) with compounds as row and FIR parameters as columns: <code>rtMin</code> (float in seconds), <code>rtMax</code> (float in seconds), <code>mzMin</code> (float), <code>mzMax</code> (float).
uROI	A data.frame of updated Regions Of Interest (uROI) with compounds as row and uROI parameters as columns: <code>rtMin</code> (float in seconds), <code>rt</code> (float in seconds, or <i>NA</i>), <code>rtMax</code> (float in seconds), <code>mzMin</code> (float), <code>mz</code> (float or <i>NA</i>), <code>mzMax</code> (float).
filepath	A character vector of file paths, of length number of spectra files
cpdMetadata	A data.frame of compound metadata, with compounds as row and metadata as columns
spectraMetadata	A data.frame of sample metadata, with samples as row and metadata as columns
acquisitionTime	A character vector of acquisition date-time (converted from POSIXct) or NA
uROIExist	A logical stating if uROI have been set
useUROI	A logical stating if uROI are to be used
useFIR	A logical stating if FIR are to be used
TIC	A numeric vector of TIC or NA, of length number of spectra files
peakTables	A list of peakTable data.frame, of length number of spectra files. Each peakTable data.frame has compounds as rows and peak annotation results as columns.
dataPoints	A list of length number of spectra files. Each list element is a <i>ROIsDataPoint</i> list of data.frame of raw data points for each ROI/uROI (retention time 'rt', mass 'mz' and intensity 'int' (as column) of each raw data points (as row))

`peakFit` A list of length number of spectra files. Each list element is a *curveFit* list of `peakPantheR_curveFit` or NA for each ROI

`isAnnotated` A logical stating if the annotation has taken place

Details:: The `peakTables` data.frame are structured as follow:

<code>cpdID</code>	database compound ID
<code>cpdName</code>	compound name
<code>found</code>	was the peak found
<code>rt</code>	retention time of peak apex (sec)
<code>rtMin</code>	leading edge of peak retention time (sec) determined at 0.5% of apex intensity
<code>rtMax</code>	trailing edge of peak retention time (sec) determined at 0.5% of apex intensity
<code>mz</code>	weighted (by intensity) mean of peak m/z across scans
<code>mzMin</code>	m/z peak minimum (between <code>rtMin</code> , <code>rtMax</code>)
<code>mzMax</code>	m/z peak maximum (between <code>rtMin</code> , <code>rtMax</code>)
<code>peakArea</code>	integrated peak area
<code>peakAreaRaw</code>	integrated peak area from raw data points
<code>maxIntMeasured</code>	maximum peak intensity in raw data
<code>maxIntPredicted</code>	maximum peak intensity based on curve fit
<code>is_filled</code>	Logical indicate if the feature was integrated using FIR (Fallback Integration Region)
<code>ppm_error</code>	difference in ppm between the expected and measured m/z
<code>rt_dev_sec</code>	difference in seconds between the expected and measured rt
<code>tailingFactor</code>	the tailing factor is a measure of peak tailing. It is defined as the distance from the front slope of the peak to the apex
<code>asymmetryFactor</code>	the asymmetry factor is a measure of peak tailing. It is defined as the distance from the center line of the peak to the apex

See Also

Other `peakPantheR`: [peakPantheR_parallelAnnotation\(\)](#), [peakPantheR_singleFileSearch\(\)](#)

Other `parallelAnnotation`: [peakPantheR_parallelAnnotation\(\)](#), [peakPantheR_singleFileSearch\(\)](#)

Examples

```
if(requireNamespace('faahKO')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahKO)
  spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                   system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                   system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

  # targetFeatTable
  targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
    c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
  targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
    522.2, 522.205222)
  targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
    496.2, 496.204962)
  targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
    FUN.VALUE=numeric(2))

  annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
```

```
targetFeatTable=targetFeatTable)
```

```

annotation
# An object of class peakPantheRAnnotation
# 2 compounds in 3 samples.
# updated ROI do not exist (uROI)
# does not use updated ROI (uROI)
# does not use fallback integration regions (FIR)
# is not annotated

slotNames(annotation)
# [1] 'cpdID'      'cpdName'    'ROI'        'FIR'        'uROI'
# [6] 'filepath'   'cpdMetadata' 'spectraMetadata' 'acquisitionTime'
# [10] 'uROIExist' 'useUROI'    'useFIR'     'TIC'        'peakTables'
# [15] 'dataPoints' 'peakFit'    'isAnnotated'

## Slots shouldn't be accessed directly, accessors are available:
cpdID(annotation)
# [1] 'ID-1' 'ID-2'
cpdName(annotation)
# [1] 'Cpd 1' 'Cpd 2'
ROI(annotation)
#   rtMin   rt   rtMax   mzMin   mz    mzMax cpdID cpdName
# 1  3310 3344.888 3390 522.1948 522.2 522.2052 ID-1  Cpd 1
# 2  3280 3385.577 3440 496.1950 496.2 496.2050 ID-2  Cpd 2
FIR(annotation)
#   rtMin rtMax mzMin mzMax cpdID cpdName
# 1   NA   NA   NA   NA  ID-1  Cpd 1
# 2   NA   NA   NA   NA  ID-2  Cpd 2
uROI(annotation)
#   rtMin rt   rtMax mzMin mz    mzMax cpdID cpdName
# 1   NA NA   NA   NA NA   NA  ID-1  Cpd 1
# 2   NA NA   NA   NA NA   NA  ID-2  Cpd 2
filepath(annotation)
# [1] 'C:/R/R-3.6.0/library/faahKO/cdf/KO/ko15.CDF'
# [2] 'C:/R/R-3.6.0/library/faahKO/cdf/KO/ko16.CDF'
# [3] 'C:/R/R-3.6.0/library/faahKO/cdf/KO/ko18.CDF'
cpdMetadata(annotation)
# data frame with 0 columns and 2 rows
spectraMetadata(annotation)
# data frame with 0 columns and 3 rows
acquisitionTime(annotation)
# [1] NA NA NA
uROIExist(annotation)
# [1] FALSE
useUROI(annotation)
# [1] FALSE
useFIR(annotation)
# [1] FALSE
TIC(annotation)
# [1] NA NA NA
peakTables(annotation)
# [[1]]
# NULL
# [[2]]
# NULL
# [[3]]

```

```

# NULL
dataPoints(annotation)
# [[1]]
# NULL
# [[2]]
# NULL
# [[3]]
# NULL
peakFit(annotation)
# [[1]]
# NULL
# [[2]]
# NULL
# [[3]]
# NULL
isAnnotated(annotation)
# [1] FALSE
}

```

peakPantheR_applyRTCorrection

Correct targeted retention time based on reference compounds

Description

Correct targeted features retention time using the RT and RT deviation of previously fitted compounds. The 'method' and 'params' are used to select and parametrise the retention time correction method employed. When 'robust' is set to TRUE, the RANSAC algorithm is used to automatically flag outliers and robustify the correction function fitting.

Usage

```

peakPantheR_applyRTCorrection(
  targetFeatTable,
  referenceTable,
  method = "polynomial",
  params = list(polynomialOrder = 3),
  robust = TRUE
)

```

Arguments

targetFeatTable a [data.frame](#) of compounds to target as rows and parameters as columns: cpdID (str), cpdName (str), rtMin (float in seconds), rt (float in seconds, or NA), rtMax (float in seconds), mzMin (float), mz (float or NA), mzMax (float).

referenceTable a [data.frame](#) of reference compound information as rows and properties as columns: cpdID (str), cpdName (str), rt (float), rt_dev_sec (float)

method (str) name of RT correction method to use (currently polynomial)

params (list) list of parameters to pass to the each correction method. Currently allowed inputs are polynomialOrder for method='polynomial'

robust (bool) whether to use the RANSAC algorithm to flag and ignore outliers during retention time correction

Value

a targetFeatTable with corrected RT

peakPanther_loadAnnotationParamsCSV
Load fit parameters from CSV

Description

Initialise a new peakPantherAnnotation object after loading ROI, uROI and FIR parameters from CSV. spectraPaths, spectraMetadata or cpdMetadata are not initialised and will need to be filled before annotation. useUROI and useFIR are set to FALSE and will need to be set accordingly. uROIExist is established depending on the uROI columns present in the CSV and will be set to TRUE only if no NA are present

Usage

```
peakPanther_loadAnnotationParamsCSV(CSVParamPath, verbose = TRUE)
```

Arguments

CSVParamPath (str) Path to a CSV file of fit parameters (e.g. as saved by outputAnnotationDiagnostic)
 verbose (bool) If TRUE message progress

Value

(peakPantherAnnotation) Object initialised with ROI, uROI and FIR read from the CSV file

Examples

```
## Input data
input_CSV <- data.frame(matrix(nrow=2, ncol=21, dimnames=list(c(),
  c('cpdID', 'cpdName',
  'X', 'ROI_rt', 'ROI_mz', 'ROI_rtMin', 'ROI_rtMax', 'ROI_mzMin', 'ROI_mzMax',
  'X', 'uROI_rtMin', 'uROI_rtMax', 'uROI_mzMin', 'uROI_mzMax', 'uROI_rt',
  'uROI_mz', 'X', 'FIR_rtMin', 'FIR_rtMax', 'FIR_mzMin', 'FIR_mzMax'))))
input_CSV[1,] <- c('ID-1', 'Cpd 1', '|', 1., 2., 3., 4., 5., 6., '|',
  7., 8., 9., 10., 11., 12., '|', 13., 14., 15., 16.)
input_CSV[2,] <- c('ID-2', 'Cpd 2', '|', 17., 18., 19., 20., 21., 22., '|',
  23., 24., 25., 26., 27., 28., '|', 29., 30., 31., 32.)
input_CSV[-c(1,2,3,10,17)] <- vapply(input_CSV[-c(1,2,3,10,17)],
  as.numeric, FUN.VALUE=numeric(2))

# temporary file location
savePath1 <- tempfile(pattern='file', tmpdir=tempdir(), fileext='.csv')
# save csv
utils::write.csv(input_CSV, file=savePath1, row.names=FALSE)

# Load parameters from CSV
```

```
loadedAnnotation <- peakPantheR_loadAnnotationParamsCSV(savePath1,
                                                       verbose=TRUE)

# uROIExist set to TRUE
# New peakPantheRAnnotation object initialised for 2 compounds
# An object of class peakPantheRAnnotation
# 2 compounds in 0 samples.
# updated ROI exist (uROI)
# does not use updated ROI (uROI)
# does not use fallback integration regions (FIR)
# is not annotated
```

```
peakPantheR_parallelAnnotation
```

Search, integrate and report targeted features in a multiple spectra

Description

Integrate all target features in all files defined in the initialised input object and store results. The use of updated ROI and the integration of FIR are controlled by the input object slots `useUROI` and `useFIR`. Files are processed in parallel using [peakPantheR_singleFileSearch](#); `ncores` controls the number of cores used for parallelisation, with `ncores=0` corresponding to serial processing. If the processing of a file fails (file does not exist or error during execution) the sample is removed from the outputted object.

Usage

```
peakPantheR_parallelAnnotation(
  object,
  ncores = 0,
  getAcquTime = TRUE,
  resetWorkers = 1,
  centroided = TRUE,
  curveModel = "skewedGaussian",
  verbose = TRUE,
  ...
)
```

Arguments

<code>object</code>	(<code>peakPantheRAnnotation</code>) Initialised <code>peakPantheRAnnotation</code> object defining the samples to process and compounds to target. The slots <code>useUROI</code> and <code>useFIR</code> controls if <code>uROI</code> must be used and <code>FIR</code> integrated if a feature is not found
<code>ncores</code>	(<code>int</code>) Number of cores to use for parallelisation. Default 0 for no parallelisation.
<code>getAcquTime</code>	(<code>bool</code>) If <code>TRUE</code> will extract sample acquisition date-time from the <code>mzML</code> meta-data (the additional file access will impact run time)
<code>resetWorkers</code>	(<code>int</code>) If 0, the parallel cluster is only initiated once. If <code>>0</code> the cluster will be reset (and the memory of each worker freed) once <code>ncores * resetWorkers</code> files have been processed. Default value is 1, the cluster is reset once <code>ncores</code> files have been processed. While potentially impacting performance (need to wait until all <code>ncores * resetWorkers</code> files are processed before restarting the cluster), shutting down the workers processes regularly will ensure the OS can reallocate

	memory more efficiently. For values >1, ensure sufficient system memory is available
centroided	(bool) use TRUE if the data is centroided, used by readMSData when reading the raw data files
curveModel	(str) specify the peak-shape model to fit, by default skewedGaussian. Accepted values are skewedGaussian and emgGaussian
verbose	(bool) If TRUE message calculation progress, time taken, number of features found (total and matched to targets) and failures
...	Passes arguments to findTargetFeatures to alter peak-picking parameters

Value

a list: `list()$result` (*peakPantheRAnnotation*) fully annotated object, `list()$failures` (*list*) list of failed samples and error message

See Also

Other peakPantheR: [peakPantheRAnnotation](#), [peakPantheR_singleFileSearch\(\)](#)

Other parallelAnnotation: [peakPantheRAnnotation](#), [peakPantheR_singleFileSearch\(\)](#)

Examples

```
if(requireNamespace('faahK0')){
  ## Load data
  library(faahK0)

  # 3 files
  input_spectraPaths <- c(system.file('cdf/K0/ko15.CDF', package = 'faahK0'),
                          system.file('cdf/K0/ko16.CDF', package = 'faahK0'),
                          system.file('cdf/K0/ko18.CDF', package = 'faahK0'))

  # 4 features
  input_ROI <- data.frame(matrix(vector(), 4, 8,
                                dimnames=list(c(), c('cpdID', 'cpdName', 'rtMin', 'rt',
                                                       'rtMax', 'mzMin', 'mz', 'mzMax'))),
                          stringsAsFactors=FALSE)
  input_ROI[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                    522.2, 522.205222)
  input_ROI[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                    496.2, 496.204962)
  input_ROI[3,] <- c('ID-3', 'Cpd 3', 3420., 3454.435, 3495., 464.195358,
                    464.2, 464.204642)
  input_ROI[4,] <- c('ID-4', 'Cpd 4', 3670., 3701.697, 3745., 536.194638,
                    536.2, 536.205362)
  input_ROI[,c(3:8)] <- vapply(input_ROI[,c(3:8)], as.numeric,
                              FUN.VALUE=numeric(4))

  # Initialise object
  initAnnotation <- peakPantheRAnnotation(spectraPaths=input_spectraPaths,
                                         targetFeatTable=input_ROI)

  # to use updated ROI:
  # uROIExist=TRUE, useUROI=TRUE, uROI=input_uROI
  # to use FallBack Integration Regions:
  # useFIR=TRUE, FIR=input_FIR
```

```
# Run serially
result_parallelAnnotation <- peakPantheR_parallelAnnotation(initAnnotation,
                                                           ncores=0,
                                                           getAcquTime=FALSE,
                                                           verbose=TRUE)

# Processing 4 compounds in 3 samples:
# uROI:\tFALSE
# FIR:\tFALSE
# ----- ko15 -----
# Polarity can not be extracted from netCDF files, please set manually the
# polarity with the 'polarity' method.
# Reading data from 4 windows
# Data read in: 0.24 secs
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #1
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #3
# Found 4/4 features in 0.06 secs
# Peak statistics done in: 0.02 secs
# Feature search done in: 0.76 secs
# ----- ko16 -----
# Polarity can not be extracted from netCDF files, please set manually the
# polarity with the 'polarity' method.
# Reading data from 4 windows
# Data read in: 0.24 secs
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #1
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #2
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #3
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #4
# Found 4/4 features in 0.08 secs
# Peak statistics done in: 0 secs
# Feature search done in: 0.71 secs
# ----- ko18 -----
# Polarity can not be extracted from netCDF files, please set manually the
# polarity with the 'polarity' method.
# Reading data from 4 windows
# Data read in: 0.25 secs
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #1
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #2
# Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
# mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
# ROI$mzMax for ROI #4
```

```

# Found 4/4 features in 0.06 secs
# Peak statistics done in: 0 secs
# Feature search done in: 0.71 secs
# -----
# Parallel annotation done in: 2.18 secs

# No failures
result_parallelAnnotation$failures

result_parallelAnnotation$annotation
# An object of class peakPantherAnnotation
# 4 compounds in 3 samples.
# updated ROI do not exist (uROI)
# does not use updated ROI (uROI)
# does not use fallback integration regions (FIR)
# is annotated
}

```

```
peakPanther_plotEICFit
```

Plot samples raw data and detected feature for a single ROI

Description

plot a ROI across multiple samples (x axis is RT, y axis is intensity). If curveFit is provided, the fitted curve for each sample is added.

Usage

```

peakPanther_plotEICFit(
  ROIDataPointSampleList,
  curveFitSampleList = NULL,
  rtMin = NULL,
  rtMax = NULL,
  sampling = 250,
  sampleColour = NULL,
  verbose = TRUE
)

```

Arguments

ROIDataPointSampleList	(list) list of data.frame of raw data points for each sample (retention time 'rt', mass 'mz' and intensity 'int' (as column) of each raw data points (as row)).
curveFitSampleList	(list) NULL or a list of peakPanther_curveFit (or NA) for each sample
rtMin	(float) NULL or vector of detected peak minimum retention time (in sec)
rtMax	(float) NULL or vector of detected peak maximum retention time (in sec)
sampling	(int) Number of points to employ when plotting fittedCurve
sampleColour	(str) NULL or vector colour for each sample (same length as ROIDataPointSampleList, rtMin, rtMax)
verbose	(bool) if TRUE message when NA scans are removed

Value

Grob (ggplot object)

Examples

```
## Input data
# fake sample 1
# ROI data points
rt1      <- seq(990, 1010, by=20/250)
mz1      <- rep(522., length(rt1))
int1     <- (dnorm(rt1, mean=1000, sd=1.5) * 100) + 1
tmp_DataPoints1 <- data.frame(rt=rt1, mz=mz1, int=int1)
# fittedCurve
fit1     <- list(amplitude=37.068916502809756, center=999.3734222573454,
                sigma=0.58493182568124724, gamma=0.090582029276037035,
                fitStatus=2, curveModel='skewedGaussian')
class(fit1) <- 'peakPantheR_curveFit'

# fake sample 2
# ROI data points
rt2      <- seq(990, 1010, by=20/250)
mz2      <- rep(522., length(rt2))
int2     <- (dnorm(rt2, mean=1002, sd=1.5) * 100) + 1
tmp_DataPoints2 <- data.frame(rt=rt2, mz=mz2, int=int2)
# fittedCurve
fit2     <- list(amplitude=37.073067416755556, center=1001.3736564832565,
                sigma=0.58496485738212201, gamma=0.090553713725151905,
                fitStatus=2, curveModel='skewedGaussian')
class(fit2) <- 'peakPantheR_curveFit'

## Plot features in 1 sample without colours
peakPantheR_plotEICFit(ROIDataPointSampleList=list(tmp_DataPoints1),
                      curveFitSampleList=list(fit1),
                      rtMin=995., rtMax=1005.,
                      sampling=250, sampleColour=NULL, verbose=FALSE)

## Plot features in 2 samples with colours
peakPantheR_plotEICFit(
  ROIDataPointSampleList=list(tmp_DataPoints1, tmp_DataPoints2),
  curveFitSampleList=list(fit1, fit2),
  rtMin=c(995., 997.), rtMax=c(1005., 1007.),
  sampling=250, sampleColour=c('blue', 'red'), verbose=FALSE)
```

peakPantheR_plotPeakwidth

Plot peak value and peakwidth by acquisition time or in input order

Description

For a single ROI, plot the peak value and peakwidth (RT, m/z, ...) of detected peaks across multiple samples, by acquisition time or in input order. If rotateAxis=FALSE x is run order / plot order, y is the apexValue / widthMin / widthMax, if rotateAxis=TRUE x is the measurement values and y the run order.


```
## Plot 4 samples with colour, rotate axis
peakPantheR_plotPeakwidth(apexValue=apexVal, widthMin=minVal,widthMax=maxVal,
                           acquTime=NULL, varName='Test variable 3',
                           sampleColour=c('blue','red','green','orange'),
                           rotateAxis=TRUE, verbose=FALSE)

## Plot 4 samples with colour by acquisition time, rotate axis
peakPantheR_plotPeakwidth(apexValue=apexVal, widthMin=minVal,widthMax=maxVal,
                           acquTime=acqTime, varName='Test variable 4',
                           sampleColour=c('blue','red','green','orange'),
                           rotateAxis=FALSE, verbose=FALSE)
```

peakPantheR_quickEIC *Extract and plot a EIC from a raw data file*

Description

Simple plot of an Extracted Ion Chromatogram (EIC) from a raw data file and a provided mz and rt window. Return ggplot plot object.

Usage

```
peakPantheR_quickEIC(
  spectraPath,
  rt,
  mz,
  valuesOnly = "Plot",
  centroided = TRUE,
  msLevel = 1L,
  verbose = TRUE
)
```

Arguments

spectraPath	(str) Path to the raw data file to read (uses 'MSnbase::readMSData()')
rt	(numeric(2) or two-column matrix) the lower and upper retention time range from which the data should be extracted. If a matrix is passed, each row corresponds to a different window. If not provided, the full retention time range will be extracted.
mz	(numeric(2) or two-column matrix) the lower and upper mass range from which the data should be extracted. If a matrix is passed, each row corresponds to a different window. If not provided, the full mass range will be extracted.
valuesOnly	(str) If 'Raw' only load the file and return a table of raw extracted values (exported version of 'extractSignalRawData()'). If 'EIC' return a table of EIC data point. Else return the EIC plot (default).
centroided	(bool) Indicate to 'MSnbase::readMSData()' whether the spectra file is centroided or not (default to 'TRUE').
msLevel	(int) The MS level at which the data should be extracted (default to MS level 1).
verbose	(bool) Output progress information or not.

Value

Grob (ggplot object) of the EIC plot, if 'valuesOnly='Raw'' returns a data.frame of raw datapoints with as columns 'rt', 'mz' and 'int'. If 'valuesOnly='EIC'' returns a data.frame of EIC datapoints with as columns 'rt' and 'int'.

Examples

```
## Use a file from the faahK0 package and plot an EIC of interest
library(faahK0)
spectraPath <- system.file('cdf/K0/ko15.CDF', package='faahK0')
peakPanther_quickEIC(spectraPath,
                     rt = c(3290., 3410.),
                     mz = c(522.194778, 522.205222))
```

```
peakPanther_ROIStatistics
```

Save to disk each ROI EIC and mean IS RT

Description

Using reference samples (referenceSpectraFiles), save (to saveFolder) each ROI EIC (ROI) and reports the mean apex RT for all IS (IS_ROI) across samples

Usage

```
peakPanther_ROIStatistics(
  referenceSpectraFiles,
  saveFolder,
  ROI = NULL,
  IS_ROI = NULL,
  sampleColour = NULL,
  ncores = 0,
  saveISPlots = TRUE,
  verbose = TRUE
)
```

Arguments

referenceSpectraFiles	(str) A character vector of paths to the reference spectra files
saveFolder	(str) Path to the folder where EICs and IS mean RT (IS_mean_RT.csv) will be saved
ROI	(data.frame) NULL or a data.frame of Regions Of Interest (ROI) with compounds as row and ROI parameters as columns: rtMin (float in seconds), rt (float in seconds, or NA), rtMax (float in seconds), mzMin (float), mz (float or NA), mzMax (float) (if NULL, ROI EICs are not saved)
IS_ROI	(data.frame) NULL or a data.frame of IS ROI with IS as row and ROI parameters as columns: rtMin (float in seconds), rt (float in seconds, or NA), rtMax (float in seconds), mzMin (float), mz (float or NA), mzMax (float) (if NULL IS mean RT is not calculated and saved in IS_mean_RT.csv)

sampleColour (str) NULL or vector colour for each sample
 ncores (int) Number of cores to use to integrate IS in parallel
 saveISPlots (bool) If TRUE save a diagnostic plot for each IS to saveFolder/IS_search compound
 verbose (bool) If TRUE message progress

Value

None

Examples

```
if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 2 samples and 1 targeted
## compound

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 1, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(1))

# input
refSpecFiles <- spectraPaths
input_ROI <- targetFeatTable
input_IS_ROI <- targetFeatTable
sampleColour <- c('blue', 'red')

# temporary saveFolder
saveFolder1 <- tempdir()

# Calculate ROI statistics
peakPantheR_ROIStatistics(refSpecFiles, saveFolder1, ROI=input_ROI,
  IS_ROI=input_IS_ROI, sampleColour=sampleColour,
  ncores=0, saveISPlots=TRUE, verbose=TRUE)
}
```

peakPantheR_singleFileSearch

Search, integrate and report targeted features in a raw spectra

Description

Report for a raw spectra the TIC, acquisition time, integrated targeted features, fitted curves and datapoints for each region of interest. Optimised to reduce the number of file access. Features not detected can be integrated using fallback integration regions (FIR).

Usage

```

peakPantheR_singleFileSearch(
  singleSpectraDataPath,
  targetFeatTable,
  peakStatistic = FALSE,
  plotEICsPath = NA,
  getAcquTime = FALSE,
  FIR = NULL,
  centroided = TRUE,
  curveModel = "skewedGaussian",
  verbose = TRUE,
  ...
)

```

Arguments

singleSpectraDataPath (str) path to netCDF or mzML raw data file (centroided, **only with the channel of interest**).

targetFeatTable a [data.frame](#) of compounds to target as rows. Columns: cpdID (str), cpdName (str), rtMin (float in seconds), rt (float in seconds, or *NA*), rtMax (float in seconds), mzMin (float), mz (float or *NA*), mzMax (float).

peakStatistic (bool) If TRUE calculates additional peak statistics: 'ppm_error', 'rt_dev_sec', 'tailing factor' and 'asymmetry factor'

plotEICsPath (str or NA) If not NA, will save a *.png* of all ROI EICs at the path provided ('filepath/filename.png' expected). If NA no plot saved

getAcquTime (bool) If TRUE will extract sample acquisition date-time from the mzML metadata (the additional file access will impact run time)

FIR (data.frame or NULL) If not NULL, integrate Fallback Integration Regions (FIR) when a feature is not found. Compounds as row are identical to **targetFeatTable**, columns are rtMin (float in seconds), rtMax (float in seconds), mzMin (float), mzMax (float).

centroided (bool) use TRUE if the data is centroided, used by [readMSData](#) when reading the raw data file

curveModel (str) specify the peak-shape model to fit, by default skewedGaussian. Accepted values are skewedGaussian and emgGaussian

verbose (bool) If TRUE message calculation progress, time taken and number of features found

... Passes arguments to [findTargetFeatures](#) to alter peak-picking parameters (e.g. curveModel, sampling, params as a list of parameters for each ROI or 'guess',...)

Value

a list: list()\$TIC (*int*) TIC value, list()\$peakTable (*data.frame*) targeted features results (see Details), list()\$curveFit (*list*) list of peakPantheR_curveFit or NA for each ROI, list()\$acquTime (*POSIXct* or *NA*) date-time of sample acquisition from mzML metadata, list()\$ROIsDataPoint (*list*) a list of data.frame of raw data points for each ROI (retention time 'rt', mass 'mz' and intensity 'int' (as column) of each raw data points (as row)).

Details:: The returned *peakTable* data.frame is structured as follow:

cpdID	database compound ID
cpdName	compound name
found	was the peak found
rt	retention time of peak apex (sec)
rtMin	leading edge of peak retention time (sec) determined at 0.5% of apex intensity
rtMax	trailing edge of peak retention time (sec) determined at 0.5% of apex intensity
mz	weighted (by intensity) mean of peak m/z across scans
mzMin	m/z peak minimum (between rtMin, rtMax)
mzMax	m/z peak maximum (between rtMin, rtMax)
peakArea	integrated peak area
peakAreaRaw	integrated peak area from raw data points
maxIntMeasured	maximum peak intensity in raw data
maxIntPredicted	maximum peak intensity based on curve fit
is_filled	Logical indicate if the feature was integrated using FIR (Fallback Integration Region)
ppm_error	difference in ppm between the expected and measured m/z
rt_dev_sec	difference in seconds between the expected and measured rt
tailingFactor	the tailing factor is a measure of peak tailing. It is defined as the distance from the front slope of the peak to the peak apex.
asymmetryFactor	the asymmetry factor is a measure of peak tailing. It is defined as the distance from the center line of the peak to the peak apex.

See Also

Other peakPanther: [peakPantherAnnotation](#), [peakPanther_parallelAnnotation\(\)](#)

Other parallelAnnotation: [peakPantherAnnotation](#), [peakPanther_parallelAnnotation\(\)](#)

Examples

```
if(requireNamespace('faahKO')){
  ## Load data
  library(faahKO)
  netcdfFilePath <- system.file('cdf/KO/ko15.CDF', package = 'faahKO')

  ## targetFeatTable
  targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
    c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
  targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
    522.2, 522.205222)
  targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
    496.2, 496.204962)
  targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
    FUN.VALUE=numeric(2))

  res <- peakPanther_singleFileSearch(netcdfFilePath, targetFeatTable,
    peakStatistic=TRUE)

  # Polarity can not be extracted from netCDF files, please set manually the
  # polarity with the 'polarity' method.
  # Reading data from 2 windows
  # Data read in: 0.16 secs
  # Warning: rtMin/rtMax outside of ROI; datapoints cannot be used for
  # mzMin/mzMax calculation, approximate mz and returning ROI$mzMin and
  # ROI$mzMax for ROI #1
  # Found 2/2 features in 0.05 secs
```

```
# Peak statistics done in: 0 secs
# Feature search done in: 0.75 secs

res
# $TIC
# [1] 2410533091
#
# $peakTable
#   found   rtMin      rt   rtMax   mzMin   mz   mzMax peakArea
# 1  TRUE 3309.759 3346.828 3385.410 522.1948 522.2 522.2052 26133727
# 2  TRUE 3345.377 3386.529 3428.279 496.2000 496.2 496.2000 35472141
#   peakAreaRaw maxIntMeasured maxIntPredicted cpdID cpdName is_filled
# 1    26071378         889280         901015.8 ID-1  Cpd 1     FALSE
# 2    36498367        1128960        1113576.7 ID-2  Cpd 2     FALSE
#   ppm_error  rt_dev_sec  tailingFactor  asymmetryFactor
# 1 0.02337616   1.9397590     1.015357         1.026824
# 2 0.02460103   0.9518072     1.005378         1.009318
#
# $acquTime
# [1] NA
#
#
# $curveFit
# $curveFit[[1]]
# $amplitude
# [1] 162404.8
#
# $center
# [1] 3341.888
#
# $sigma
# [1] 0.07878613
#
# $gamma
# [1] 0.00183361
#
# $fitStatus
# [1] 2
#
# $curveModel
# [1] 'skewedGaussian'
#
# attr(,"class")
# [1] 'peakPantheR_curveFit'
#
# $curveFit[[2]]
# $amplitude
# [1] 199249.1
#
# $center
# [1] 3382.577
#
# $sigma
# [1] 0.07490442
#
# $gamma
# [1] 0.00114719
```

```

#
# $fitStatus
# [1] 2
#
# $curveModel
# [1] 'skewedGaussian'
#
# attr(,"class")
# [1] 'peakPantheR_curveFit'
#
#
# $ROIsDataPoint
# $ROIsDataPoint[[1]]
#      rt    mz    int
# 1 3315.154 522.2  2187
# 2 3316.719 522.2  3534
# 3 3318.284 522.2  6338
# 4 3319.849 522.2 11718
# 5 3321.414 522.2 21744
# 6 3322.979 522.2 37872
# 7 3324.544 522.2 62424
# 8 3326.109 522.2 98408
# 9 3327.673 522.2 152896
# 10 3329.238 522.2 225984
# ...
#
# $ROIsDataPoint[[2]]
#      rt    mz    int
# 1 3280.725 496.2  1349
# 2 3290.115 496.2  2069
# 3 3291.680 496.2  3103
# 4 3293.245 496.2  5570
# 5 3294.809 496.2 10730
# 6 3296.374 496.2 20904
# 7 3297.939 496.2 38712
# 8 3299.504 496.2 64368
# 9 3301.069 496.2 97096
# 10 3302.634 496.2 136320
# ...
}

```

peakPantheR_start_GUI *peakPantheR Graphical User Interface*

Description

peakPantheR Graphical User Interface (GUI) implements all the functions for the parallel detection, integration and reporting of pre-defined features in multiple mass spectrometry data files. To exit press ESC in the command line.

Usage

```
peakPantheR_start_GUI(browser = TRUE)
```

Arguments

browser If TRUE open the graphical user interface in a web browser instead of a R window. Default is TRUE

Value

None, start GUI. To exit press ESC in the command line.

Examples

```
print("Start graphical interface, press 'ESC' in the command line to stop")
# peakPantheR_start_GUI()
```

peakTables, peakPantheRAnnotation-method

peakTables accessor with cpdID and cpdName added back

Description

peakTables accessor with cpdID and cpdName added back

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
peakTables(object)
```

Arguments

object peakPantheRAnnotation

Value

(data.frame) A list of peakTable data.frame, of length number of spectra files. Each peakTable data.frame has compounds as rows and peak annotation results as columns, with added compound ID and name.

Examples

```
if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
'mzMax'))), stringsAsFactors=FALSE)
```

```

targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                        522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                        496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
                                  FUN.VALUE=numeric(2))

annotation <- peakPantherAnnotation(spectraPaths=spectraPaths,
                                   targetFeatTable=targetFeatTable)

## default values without annotation
peakTables(annotation)
# [[1]]
# NULL
# [[2]]
# NULL
# [[3]]
# NULL
}

```

plotEICDetectedPeakwidth

Plot samples raw data and detected feature for a single ROI

Description

Plot a ROI across multiple samples (x axis is RT, y axis is intensity) with the matching detected peak rt and peakwidth under it. If curveFit is provided, the fitted curve for each compound is added. RT and peakwidth are plotted in the order spectra are passed, with the first spectra on top.

Usage

```

plotEICDetectedPeakwidth(
  ROIDataPointSampleList,
  cpdID,
  cpdName,
  rt,
  rtMin,
  rtMax,
  mzMin,
  mzMax,
  ratio = 0.85,
  sampling = 250,
  curveFitSampleList = NULL,
  sampleColour = NULL,
  verbose = TRUE
)

```

Arguments

ROIDataPointSampleList

(list) list of data.frame of raw data points for each sample (retention time 'rt', mass 'mz' and intensity 'int' (as column) of each raw data points (as row)).

cpdID	(str) Compound ID
cpdName	(str) Compound Name
rt	(float) vector of detected peak apex retention time (in sec)
rtMin	(float) vector of detected peak minimum retention time (in sec)
rtMax	(float) vector of detected peak maximum retention time (in sec)
mzMin	(float) ROI minimum m/z (matching EIC)
mzMax	(float) ROI maximum m/z (matching EIC)
ratio	(float) value between 0 and 1 defining the vertical percentage taken by the EICs subplot
sampling	(int) Number of points to employ when plotting fittedCurve
curveFitSampleList	(list) NULL or a list of peakPanther_curveFit (or NA) for each sample
sampleColour	(str) NULL or vector colour for each sample (same length as EICs, rt, rtMin, rtMax)
verbose	(bool) if TRUE message when NA scans are removed

Value

Grob (ggplot object)

plotHistogram	<i>Plot variable histogram and density</i>
---------------	--

Description

Plot the histogram and density of the variable

Usage

```
plotHistogram(var, varName = "Variable", density = TRUE, ...)
```

Arguments

var	(float) vector of values to plot
varName	(str) Name of the variable to plot
density	(bool) If TRUE plot overlay the density on the variable
...	Passes arguments to ggplot2::geom_histogram, e.g. bins=20, binwidth=1

Value

Grob (ggplot object)

predictCurve *Predict curve values*

Description

Evaluate fitted curve values at x data points

Usage

```
predictCurve(fittedCurve, x)
```

Arguments

fittedCurve (peakPantheR_curveFit) A 'peakPantheR_curveFit': a list of curve fitting parameters, curve shape model curveModel and nls.lm fit status fitStatus.

x (numeric) values at which to evaluate the fitted curve

Details

```
## Examples cannot be computed as the function is not exported: ## Input a fitted curve fittedCurve
<- list(amplitude=275371.1, center=3382.577, sigma=0.07904697, gamma=0.001147647, fitStatus=2, curveModel='skewedGaussian') class(fittedCurve) <- 'peakPantheR_curveFit' input_x <-
c(3290, 3300, 3310, 3320, 3330, 3340, 3350, 3360, 3370, 3380, 3390, 3400, 3410)

## Predict y at each input_x pred_y <- predictCurve(fittedCurve, input_x) pred_y # [1] 2.347729e-
08 1.282668e-05 3.475590e-03 4.676579e-01 3.129420e+01 # [6] 1.043341e+03 1.736915e+04
1.447754e+05 6.061808e+05 1.280037e+06 # [11] 1.369651e+06 7.467333e+05 2.087477e+05
```

Value

fitted curve values at x

prepare_advanced_target_parameters
*Process target region parameters (with uROI, FIR) for object initiali-
sation*

Description

Process target region parameters with uROI and FIR (cpdID, cpdName, ROI_rt, ROI_mz, ROI_rtMin, ROI_rtMax, ROI_mzMin, ROI_mzMax, uROI_rtMin, uROI_rtMax, uROI_mzMin, uROI_mzMax, uROI_rt, uROI_mz, FIR_rtMin, FIR_rtMax, FIR_mzMin, FIR_mzMax) and return input variables for peakPantheRAnnotation()

Usage

```
prepare_advanced_target_parameters(paramTable, verbose)
```

Arguments

paramTable (data.frame) Target region parameters

verbose (bool) If TRUE message progress

Value

(list) List of targetFeatTable, uROI, FIR, uROIExist

prepare_basic_target_parameters

Process target region parameters (simple format) for object initialisation

Description

Process the simple target region parameters (cpdID, cpdName, mzMin, mzMax, mz, rtMin, rtMax, rt) and return input variables for peakPantheRAnnotation()

Usage

```
prepare_basic_target_parameters(paramTable)
```

Arguments

paramTable (data.frame) Target region parameters

Value

(list) List of targetFeatTable, uROI, FIR, uROIExist

resetAnnotation, peakPantheRAnnotation-method

Reset a peakPantheRAnnotation and alter samples and compounds information

Description

Reset a peakPantheRAnnotation (remove results and set isAnnotated=FALSE). If a different number of samples (spectraPaths) or compounds (targetFeatTable) are passed, the object will be initialised to the new size. For input values left as NULL, the slots (filepath (from spectraPaths), ROI, cpdID, cpdName (from targetFeatTable), uROI, FIR, cpdMetadata, spectraMetadata, uROIExist, useUROI and useFIR) will be filled with values from previousAnnotation.

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
resetAnnotation(
  previousAnnotation,
  spectraPaths = NULL,
  targetFeatTable = NULL,
  uROI = NULL,
  FIR = NULL,
  cpdMetadata = NULL,
  spectraMetadata = NULL,
```

```

    uROIExist = NULL,
    useUROI = NULL,
    useFIR = NULL,
    verbose = TRUE,
    ...
  )

```

Arguments

previousAnnotation	(peakPantheRAnnotation) object to reset
spectraPaths	NULL or a character vector of spectra file paths, to set samples to process
targetFeatTable	NULL or a <code>data.frame</code> of compounds to target as rows and parameters as columns: cpdID (str), cpdName (str), rtMin (float in seconds), rt (float in seconds, or <i>NA</i>), rtMax (float in seconds), mzMin (float), mz (float or <i>NA</i>), mzMax (float). Set compounds to target.
uROI	NULL or a <code>data.frame</code> of updated Regions Of Interest (uROI) with compounds as row and uROI parameters as columns: rtMin (float in seconds), rt (float in seconds, or <i>NA</i>), rtMax (float in seconds), mzMin (float), mz (float or <i>NA</i>), mzMax (float).
FIR	NULL or a <code>data.frame</code> of Fallback Integration Regions (FIR) with compounds as row and FIR parameters as columns: rtMin (float in seconds), rtMax (float in seconds), mzMin (float), mzMax (float).
cpdMetadata	NULL or a <code>data.frame</code> of compound metadata, with compounds as row and metadata as columns
spectraMetadata	NULL or a <code>data.frame</code> of sample metadata, with samples as row and metadata as columns
uROIExist	NULL or a logical stating if uROI have been set
useUROI	NULL or a logical stating if uROI are to be used
useFIR	NULL or a logical stating if FIR are to be used
verbose	(bool) If TRUE message progress
...	Additional slots and values to set when resetting the object (cpdID, cpdName, ROI, filepath, TIC, acquisitionTime, peakTables, dataPoints, peakFit)

Value

(peakPantheRAnnotation) object reset with previous results removed and slots updated

Examples

```

if(requireNamespace('faahK0')){
  ## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
  ## compounds

  # Paths to spectra files
  library(faahK0)
  spectraPaths <- c(system.file('cdf/K0/ko15.CDF', package = 'faahK0'),
                    system.file('cdf/K0/ko16.CDF', package = 'faahK0'))

```

```

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

smallAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

smallAnnotation
# An object of class peakPantheRAnnotation
# 2 compounds in 2 samples.
# updated ROI do not exist (uROI)
# does not use updated ROI (uROI)
# does not use fallback integration regions (FIR)
# is not annotated

# Reset and change number of spectra
newSpectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))
largerAnnotation <- resetAnnotation(smallAnnotation,
  spectraPaths=newSpectraPaths,
  verbose=TRUE)

largerAnnotation
# An object of class peakPantheRAnnotation
# 2 compounds in 3 samples.
# updated ROI do not exist (uROI)
# does not use updated ROI (uROI)
# does not use fallback integration regions (FIR)
# is not annotated
}

```

```
resetFIR,peakPantheRAnnotation-method
```

*Reset FIR windows to uROI or ROI values Reset FIR windows to uROI
(or ROI if uROIExist=FALSE)*

Description

Reset FIR windows to uROI or ROI values Reset FIR windows to uROI (or ROI if uROIExist=FALSE)

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
resetFIR(object, verbose)
```

Arguments

object	(peakPantheRAnnotation) object for which FIR are to be reset
verbose	(bool) If TRUE message progress

Value

(peakPantheRAnnotation) object with FIR values reset

Examples

```
## Initialise a peakPantheRAnnotation object with 2 targeted compounds

## targetFeatTable
input_targetFeatTable <- data.frame(matrix(vector(), 2, 8,
      dimnames=list(c(), c('cpdID', 'cpdName', 'rtMin',
        'rt', 'rtMax', 'mzMin', 'mz', 'mzMax'))),
      stringsAsFactors=FALSE)
input_targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3., 1., 4., 5., 2., 6.)
input_targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 19., 17., 20., 21., 18., 22.)
input_targetFeatTable[,c(3:8)] <- sapply(input_targetFeatTable[,c(3:8)],
      as.numeric)

## FIR
input_FIR <- data.frame(matrix(vector(), 2, 4, dimnames=list(c(),
      c('rtMin', 'rtMax', 'mzMin', 'mzMax'))),
      stringsAsFactors=FALSE)
input_FIR[1,] <- c(13., 14., 15., 16.)
input_FIR[2,] <- c(29., 30., 31., 32.)

annotation <- peakPantheRAnnotation(targetFeatTable = input_targetFeatTable,
      FIR = input_FIR, uROIExist = FALSE)

## Reset FIR with ROI values as uROI are not set
updatedAnnotation <- resetFIR(annotation, verbose=TRUE)
# FIR will be reset with ROI values as uROI values are not set
```

retentionTimeCorrection,peakPantheRAnnotation-method

Apply retention time correction methods to adjust the retention time information in the uROI of peakPantheRAnnotation object

Description

Performs retention time correction to re-adjust the expected retention time position of compounds. Requires an annotated peakPantheRAnnotation object (isAnnotated=TRUE). The original rt value is used as expected and the observed deviation measured in the rt_dev_sec field is taken as the deviation to be corrected.

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
retentionTimeCorrection(
  annotationObject,
  rtCorrectionReferences = NULL,
  method = "polynomial",
  params = list(polynomialOrder = 2),
  robust = FALSE,
  rtWindowWidth = 15,
  diagnostic = TRUE
)
```

Arguments

annotationObject	(peakPantheRAnnotation) object with previous fit results to adjust retention time values in uROI and FIR annotationObject, rtCorrectionReferences=NULL,
rtCorrectionReferences	(list) of compounds IDs (cpdID) to be used as retention time references. All cpdID entries must be present in the object and previously annotated. If NULL, use all compounds.
method	(str) name of RT correction method to use (currently polynomial or constant
params	(list) list of parameters to pass to each correction method. Currently allowed inputs are polynomialOrder for method='polynomial'
robust	(bool) whether to use the RANSAC algorithm to flag and ignore outliers during retention time correction
rtWindowWidth	(numeric) full width in seconds of the retention time window defined around the corrected retention time value for each compound
diagnostic	(bool) If TRUE returns diagnostic plots (specific to each correction method)

Value

(list) containing entries 'annotation', with the new and retention time corrected peakPantheRAnnotation, and 'plot' (if diagnostic=TRUE).

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

smallAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# annotate files serially
annotation_result <- peakPantheR_parallelAnnotation(smallAnnotation,
  ncores=0, verbose=TRUE)
data_annotation <- annotation_result$annotation

# Example with constant correction
rtCorrectionOutput <- retentionTimeCorrection(

```

```

        annotationObject = data_annotation,
        rtCorrectionReferences=c('ID-1'),
        method='constant', params=list(),
        robust=FALSE,
        rtWindowWidth=15,
        diagnostic=TRUE)

rtCorrectedAnnotation <- rtCorrectionOutput$annotation

# rtCorrectedAnnotation
# An object of class peakPantheRAnnotation
# 2 compounds in 2 samples.
# updated ROI exists, with a modified rt (uROI)
# uses updated ROI (uROI)
# uses fallback integration regions (FIR)
# is annotated

rtCorrectionPlot <- rtCorrectionOutput$plot
# rtCorrectedPlot
# A ggplot2 object
# Scatterplot where x=`r` in the and y=`rt_dev_sec` from data_annotation
# Points colored depending on whether the reference was used to fit
# the correction model
}

```

ROI,peakPantheRAnnotation-method

ROI accessor returns targetFeatTable with cpdID, cpdName added

Description

ROI accessor returns targetFeatTable with cpdID, cpdName added

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
ROI(object)
```

Arguments

object peakPantheRAnnotation

Value

(data.frame) target feature table with compounds as row and ROI parameters as columns

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)

```



```

spectraPaths <- c(system.file('cdf/K0/ko15.CDF', package = 'faahK0'),
                  system.file('cdf/K0/ko16.CDF', package = 'faahK0'),
                  system.file('cdf/K0/ko18.CDF', package = 'faahK0'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

annotation <- peakPantherAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

ROI(annotation)
#   rtMin      rt rtMax   mzMin   mz   mzMax cpdID cpdName
# 1  3310 3344.888  3390 522.1948 522.2 522.2052  ID-1   Cpd 1
# 2  3280 3385.577  3440 496.1950 496.2 496.2050  ID-2   Cpd 2
}

```

```
saveSingleFileMultiEIC
```

Save to disk a plot of all ROI EIC and detected feature range

Description

Plot and save a .png of all ROI (x is RT, y is intensity), with the matching detected peak rt and peakwidth under it.

Usage

```

saveSingleFileMultiEIC(
  ROIsDataPoint,
  curveFit,
  foundPeakTable,
  savePath,
  width = 15,
  height = 15,
  verbose = TRUE
)

```

Arguments

ROIsDataPoint (list) a list of data.frame of raw data points for each ROI (retention time 'rt', mass 'mz' and intensity 'int' (as column) of each raw data points (as row)).

curveFit (list) a list of peakPantherR_curveFit or NA for each ROI

foundPeakTable (data.frame) data.frame as generated by [findTargetFeatures](#), with features as rows and peak properties as columns. The following columns are mandatory: cpdID, cpdName, rt, rtmin, rtmax, mzmin, mzmax.

savePath	(str) Full path to save a <i>.png</i> of all ROI EICs, expect 'filepath/filename.png'.
width	(float) Width in cm for a single ROI plot (if more than one plot in total, 2 columns will be used). dpi set to a 100.
height	(float) height in a cm for a single ROI plot. dpi set to 100
verbose	(bool) if TRUE message progress

Value

None

skewedGaussian_guess *Guess function for initial skewed gaussian parameters and bounds*

Description

Guess function for initial skewed gaussian parameters and bounds, at the moment only checks the x position

Usage

```
skewedGaussian_guess(x, y)
```

Arguments

x	(numeric) x values (e.g. retention time)
y	(numeric) y observed values (e.g. spectra intensity)

Value

A list of guessed starting parameters `list()$init_params`, lower `list()$lower_bounds` and upper bounds `list()$upper_bounds` (`$gamma`, `$center`, `$sigma`, `$amplitude`)

skewedGaussian_minpack.lm
Implementation of the Skewed Gaussian peak shape for use with minpack.lm

Description

Implementation of the Skewed Gaussian peak shape for use with minpack.lm

Usage

```
skewedGaussian_minpack.lm(params, xx)
```

Arguments

params	(list) skewed gaussian parameters (<code>params\$gamma</code> , <code>params\$center</code> , <code>params\$sigma</code> , <code>params\$amplitude</code>)
xx	(numeric) values at which to evaluate the skewed gaussian

Value

value of the skewed gaussian evaluated at xx

skewedGaussian_minpack.lm_objectiveFun
Skewed Gaussian minpack.lm objective function

Description

Skewed Gaussian minpack.lm objective function, calculates residuals using the skewed gaussian Peak Shape

Usage

```
skewedGaussian_minpack.lm_objectiveFun(params, observed, xx)
```

Arguments

params	(list) skewed gaussian parameters (params\$gamma, params\$center, params\$sigma, params\$amplitude)
observed	(numeric) observed y value at xx
xx	(numeric) value at which to evaluate the skewed gaussian

Value

difference between observed and expected skewed gaussian value evaluated at xx

spectraMetadata, peakPantheRAnnotation-method
spectraMetadata accessor

Description

spectraMetadata accessor

Usage

```
## S4 method for signature 'peakPantheRAnnotation'  
spectraMetadata(object)
```

Arguments

object	peakPantheRAnnotation
--------	-----------------------

Value

(data.frame) A data.frame of sample metadata, with samples as row and metadata as columns

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantherAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
                        c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
                          'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                        522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                        496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
                                  FUN.VALUE=numeric(2))

annotation <- peakPantherAnnotation(spectraPaths=spectraPaths,
                                   targetFeatTable=targetFeatTable)

## default values not initialised
spectraMetadata(annotation)
# data frame with 0 columns and 3 rows
}

```

spectraPaths_and_metadata_UI_helper

UI data import helper - prepare file paths and metadata

Description

Return spectraPaths and spectraMetadata from a .csv file (if available). If reading from the spectraMetadata file, the spectraPaths are taken from the 'filepath' column

Usage

```

spectraPaths_and_metadata_UI_helper(
  spectraPaths = NULL,
  spectraMetadataPath = NULL
)

```

Arguments

spectraPaths NULL or character vector of spectra file paths, to set samples to process

spectraMetadataPath NULL or path to a csv of spectra metadata, with spectra as row and metadata as columns. (spectraPaths in column 'filepath')

Value

spectraPaths (str) and spectraMetadata (DataFrame or NULL) read from the CSV file

Examples

```
## Input data
# spectraPath
input_spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# spectraMetadata
input_spectraMetadata <- data.frame(matrix(data=c(input_spectraPaths,
          c('a','b','c')), nrow=3, ncol=2,
          dimnames=list(c(),c('filepath', 'testcol')),
          byrow=FALSE), stringsAsFactors=FALSE)

# temporary file location
spectraMetaPath <- tempfile(pattern="file", tmpdir=tempdir(), fileext='.csv')
# save csv
utils::write.csv(input_spectraMetadata,
                 file=spectraMetaPath,
                 row.names=FALSE)

# load data from CSV
spectraPaths_and_metadata_UI_helper(spectraPaths = NULL,
                                     spectraMetadataPath = spectraMetaPath)
```

spectra_metadata_colourScheme_UI_helper
UI export plot helper - sample colour

Description

Return a vector of spectra colours based on a metadata column

Usage

```
spectra_metadata_colourScheme_UI_helper(
  annot,
  splColrColumn = NULL
)
```

Arguments

annot (peakPantheRAnnotation) Annotation object
 splColrColumn (str) NULL, None or a spectraMetadata column for colouring each sample

Value

(character) Vector of colours

Examples

```
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
spectraPaths <- c('./path/file1', './path/file2', './path/file3')

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

emptyAnnotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

# colour scheme with no spectraMetadata
outputAnnotationFeatureMetadata_UI_helper(emptyAnnotation)
# NULL
```

TIC,peakPantheRAnnotation-method
TIC accessor

Description

TIC accessor

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
TIC(object)
```

Arguments

object peakPantheRAnnotation

Value

(float) A numeric vector of Total Ion Chromatogram or NA, of length number of spectra files

Examples

```
if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
```

```

library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
                  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

## default values without annotation
TIC(annotation)
# [1] NA NA NA
}

```

uROI,peakPantheRAnnotation-method

uROI accessor returns targetFeatTable with cpdID, cpdName added

Description

uROI accessor returns targetFeatTable with cpdID, cpdName added

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
uROI(object)
```

Arguments

object peakPantheRAnnotation

Value

(data.frame) target feature table with compounds as row and uROI parameters as columns

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),

```

```

system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

## default values without annotation
uROI(annotation)
#   rtMin rt  rtMax mzMin mz  mzMax cpdID cpdName
# 1   NA NA   NA   NA NA   NA ID-1   Cpd 1
# 2   NA NA   NA   NA NA   NA ID-2   Cpd 2
}

```

uROIExist,peakPantheRAnnotation-method

uROIExist accessor

Description

uROIExist accessor

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
uROIExist(object)
```

Arguments

object peakPantheRAnnotation

Value

(bool) flag if uROI have been set

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),

```



```

system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
  'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

uROIExist(annotation)
# [1] FALSE
}

```

useFIR,peakPantheRAnnotation-method
useFIR accessor

Description

useFIR accessor

Usage

```
## S4 method for signature 'peakPantheRAnnotation'
useFIR(object)
```

Arguments

object peakPantheRAnnotation

Value

(bool) flag if FIR are to be used

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))
}

```

```

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',
    'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
  522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
  496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
  FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
  targetFeatTable=targetFeatTable)

useFIR(annotation)
# [1] FALSE
}

```

```

useUROI,peakPantheRAnnotation-method
  useUROI accessor

```

Description

useUROI accessor

Usage

```

## S4 method for signature 'peakPantheRAnnotation'
useUROI(object)

```

Arguments

object peakPantheRAnnotation

Value

(bool) flag if uROI are to be used

Examples

```

if(requireNamespace('faahKO')){
## Initialise a peakPantheRAnnotation object with 3 samples and 2 targeted
## compounds

# Paths to spectra files
library(faahKO)
spectraPaths <- c(system.file('cdf/KO/ko15.CDF', package = 'faahKO'),
  system.file('cdf/KO/ko16.CDF', package = 'faahKO'),
  system.file('cdf/KO/ko18.CDF', package = 'faahKO'))

# targetFeatTable
targetFeatTable <- data.frame(matrix(vector(), 2, 8, dimnames=list(c(),
  c('cpdID', 'cpdName', 'rtMin', 'rt', 'rtMax', 'mzMin', 'mz',

```

```
      'mzMax'))), stringsAsFactors=FALSE)
targetFeatTable[1,] <- c('ID-1', 'Cpd 1', 3310., 3344.888, 3390., 522.194778,
                        522.2, 522.205222)
targetFeatTable[2,] <- c('ID-2', 'Cpd 2', 3280., 3385.577, 3440., 496.195038,
                        496.2, 496.204962)
targetFeatTable[,c(3:8)] <- vapply(targetFeatTable[,c(3:8)], as.numeric,
                                   FUN.VALUE=numeric(2))

annotation <- peakPantheRAnnotation(spectraPaths=spectraPaths,
                                   targetFeatTable=targetFeatTable)

useUROI(annotation)
# [1] FALSE
}
```

[,peakPantheRAnnotation,ANY,ANY,ANY-method

extract parts of peakPantheRAnnotation class

Description

extract parts of peakPantheRAnnotation class

Usage

```
## S4 method for signature 'peakPantheRAnnotation,ANY,ANY,ANY'
x[i, j, drop = "missing"]
```

Arguments

x	object from which to extract element(s) or in which to replace element(s).
i	(sample) indices specifying elements to extract or replace
j	(compound) indices specifying elements to extract or replace
drop	not applicable

Value

(peakPantheRAnnotation) object subset

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