

Package ‘ZygotyPredictor’

November 28, 2024

Type Package

Title Package for prediction of zygoty for variants/genes in NGS data

Version 1.7.0

Date 2023-12-08

Imports GenomicAlignments, GenomicRanges, Rsamtools, IRanges, VariantAnnotation, DelayedArray, dplyr, stringr, purrr, tibble, methods, knitr, igraph, readr, stats, magrittr, rlang

License GPL-2

Description

The ZygotyPredictor allows to predict how many copies of a gene are affected by small variants. In addition to the basic calculations of the affected copy number of a variant, the ZygotyPredictor can integrate the influence of several variants on a gene and ultimately make a statement if and how many wild-type copies of the gene are left. This information proves to be of particular use in the context of translational medicine. For example, in cancer genomes, the ZygotyPredictor can address whether unmutated copies of tumor-suppressor genes are present. Beyond this, it is possible to make this statement for all genes of an organism. The ZygotyPredictor was primarily developed to handle SNVs and INDELS (later addressed as small-variants) of somatic and germline origin. In order not to overlook severe effects outside of the small-variant context, it has been extended with the assessment of large scale deletions, which cause losses of whole genes or parts of them.

RoxygenNote 7.2.3

Encoding UTF-8

biocViews BiomedicalInformatics, FunctionalPrediction, SomaticMutation, GenePrediction

Depends R (>= 4.3.0)

LazyData false

Suggests rmarkdown, testthat, BiocStyle

VignetteBuilder knitr

git_url <https://git.bioconductor.org/packages/ZygotyPredictor>

git_branch devel

git_last_commit 1c2d5b8

git_last_commit_date 2024-10-29

Repository Bioconductor 3.21

Date/Publication 2024-11-27

Author Marco Rheinnecker [aut, cre] (ORCID:
<https://orcid.org/0009-0009-7181-3977>),
 Marc Ruebsam [aut],
 Daniel Huebschmann [aut],
 Martina Froehlich [aut],
 Barbara Hutter [aut]

Maintainer Marco Rheinnecker <marco.rheinnecker@dkfz-heidelberg.de>

Contents

aff_germ_copies	2
aff_som_copies	3
gene_ov	4
GR_GENE_MODEL	5
GR_GERM_SMALL_VARS	6
GR_HAPLOBLOCKS	6
GR_SCNA	7
GR_SOM_SMALL_VARS	7
predict_per_variant	8
predict_zygosity	10
ZP_ov	14
Index	16

aff_germ_copies	<i>calculates how many copies are affected by a germline small variant</i>
-----------------	--

Description

calculates how many copies are affected by a germline small variant

Usage

```
aff_germ_copies(chr, af, tcn, purity, sex, c_normal = NULL, af_normal = 0.5)
```

Arguments

chr	chromosome of the variant (either format 1,2,...,X,Y or chr1,...,chrX)
af	Allele-frequency of the variant (numeric value between 0 and 1)
tcn	total-copynumber at position of the variant (numeric value >0)

purity	purity of the sample (numeric value between 0 and 1 indicating the fraction of relevant sample with control/unrelevant tissue)
sex	sex of the sample (character: "male", "female", "m", "f")
c_normal	expected copy number at position of the variant in normal tissue, 1 for gonosomes in male samples, and 2 for male autosomes and all chromosomes in female samples. (The function can also assess the c_normal parameter by itself, but then the following two inputs must be provided: chr and sex)
af_normal	Allele-frequency in normal tissue (numeric value between 0 and 1) 0.5 represents heterozygous variants in diploid genome, 1 would be homozygous. Could be relevant if germline CNVs are present at the position. Then also the c_normal parameter would have to be adjusted.

Value

A numeric value indicating the affecting copies for the variant

Examples

```
library(dplyr)
library(purrr)
library(stringr)
aff_germ_copies(af=0.67, tcn=2, purity=0.9, chr="chrX", sex="female")
```

aff_som_copies	<i>calculates how many copies are affected by a somatic small variant</i>
----------------	---

Description

calculates how many copies are affected by a somatic small variant

Usage

```
aff_som_copies(chr, af, tcn, purity, sex, c_normal = NULL)
```

Arguments

chr	chromosome of the variant (either format 1,2,...,X,Y or chr1,...,chrX)
af	Allele-frequency of the variant (numeric value between 0 and 1)
tcn	total-copynumber at position of the variant (numeric value >0)
purity	purity of the sample (numeric value between 0 and 1 indicating the fraction of relevant sample with control/unrelevant tissue)
sex	sex of the sample (character: "male", "female", "m", "f")
c_normal	expected copy number at the position of the variant in normal tissue, 1 for gonosomes in male samples, and 2 for male autosomes and all chromosomes in female samples. (The function can also assess the c_normal parameter by itself, but then the following two inputs must be provided: chr and sex)

Value

A numeric value indicating the affecting copies for the variant

Examples

```
library(dplyr)
library(purrr)
library(stringr)
aff_som_copies(chr="chrX", af=0.67, tcn=2, purity=0.9, sex="female")
```

gene_ov	<i>accessor for gene predictions printing detailed info about how a gene status was assigned</i>
---------	--

Description

accessor for gene predictions printing detailed info about how a gene status was assigned

Usage

```
gene_ov(fp, inp_gene, n = 20)
```

Arguments

fp	full prediction (output of predict_zygoisty())
inp_gene	name of gene that should be printed with detailed information
n	max number of rows to print, as some gene status depend on loads of phasing results#

Value

prints overview about run from function predict_zygoisty() with specific information about provided gene

Examples

```
cnvs = GenomicRanges::GRanges(
  dplyr::tibble(
    chr = "chr17",
    start = c(170060, 34520990),
    end = c(34520990, 83198614),
    tcn = c(2, 1),
    cna_type = c("neutral", "LOH")
  )
)
somatic_vars = GenomicRanges::GRanges(
  dplyr::tibble(
    chr="chr17",
```

```
      start = 7675088,
      end = 7675088,
      ref = "C",
      alt = "T",
      af = 0.65,
      gene = "TP53"
    )
  )
  germline_vars = GenomicRanges::GRanges(
    dplyr::tibble(
      chr="chr17",
      start = 41771694,
      end = 41771694,
      ref = "GTGT",
      alt = "G",
      af = 0.95,
      gene = "JUP"
    )
  )
  reference = GenomicRanges::GRanges(
    dplyr::tibble(
      chr = "chr17",
      start = c(7661778, 41754603),
      end = c(7687538, 41786931),
      gene = c("TP53", "JUP")
    )
  )
  sex = "female"
  purity = 0.9
  bamfile <- system.file("extdata", "ZP_example.bam",
    package = "ZygotyPredictor")
  fp <- predict_zygoty(purity = purity, sex = sex,
    somCna = cnvs,
    somSmallVars = somatic_vars,
    germSmallVars = germline_vars,
    geneModel = reference,
    bamDna = bamfile
  )
  gene_ov(fp, TP53)
```

GR_GENE_MODEL

germline small variant object

Description

germline small variant object

Usage

data(GR_GENE_MODEL)

Format

```
## 'GR_GENE_MODEL' GRanges object
```

Value

Object containing gene model of hg38

GR_GERM_SMALL_VARS	<i>germline small variant object</i>
--------------------	--------------------------------------

Description

germline small variant object

Usage

```
data(GR_SOM_SMALL_VARS)
```

Format

```
## 'GR_SOM_SMALL_VARS' GRanges object
```

Value

Object containing germline Indels and SNVs of SeqC2 example case

GR_HAPLOBLOCKS	<i>haploblocks</i>
----------------	--------------------

Description

haploblocks

Usage

```
data(GR_HAPLOBLOCKS)
```

Format

```
## 'GR_HAPLOBLOCKS' GRanges object
```

Value

Object containing haploblock annotations

GR_SCNA	<i>copynumber object</i>
---------	--------------------------

Description

copynumber object

Usage

data(GR_SCNA)

Format

'GR_SCNA' GRanges object

Value

Object containing somatic copy number aberrations (sCNAs) of SeqC2 example case

GR_SOM_SMALL_VARS	<i>somatic small variant object</i>
-------------------	-------------------------------------

Description

somatic small variant object

Usage

data(GR_GERM_SMALL_VARS)

Format

'GR_GERM_SMALL_VARS' GRanges object

Value

Object containing somatic Indels and SNVs of SeqC2 example case

predict_per_variant *predicts zygosity of a set of variants*

Description

predicts zygosity of a set of variants

Usage

```
predict_per_variant(
  purity,
  sex,
  somCna,
  geneModel = NULL,
  somSmallVars = NULL,
  germSmallVars = NULL,
  ploidy = NULL,
  colnameTcn = NULL,
  colnameCnaType = NULL,
  includeHomoDel = TRUE,
  includeIncompleteDel = TRUE,
  assumeSomCnaGaps = FALSE,
  byTcn = TRUE,
  ZP_env = NULL,
  verbose = FALSE
)
```

Arguments

purity	purity of the sample (numeric value between 0 and 1 indicating the fraction of relevant sample with control/unrelevant tissue)
sex	sex of the sample (character: "male", "female", "m", "f")
somCna	GRanges object containing all genomic regions with annotated total copynumber and cna_type as metadata columns. The total-copynumber column should be named "tcn" but also some other commonly used names. It should contain numeric values or characters that can be converted to numeric values. The cna_type column must contain the information about loss of heterozygosity (LOH). Therefore the term "LOH" must be explicitly mentioned in the column. If a genomic region is not present in the object, it will be taken as heterozygous with neutral TCN of 2.
geneModel	GRanges object containing the gene-annotation of the used reference genome with metadata column of the gene name (gene)
somSmallVars	GRanges object containing all somatic small variants (SNV and INDEL). Required metadata columns are reference base (ref/REF), alternative base (alt/ALT), annotation of the gene name (gene/GENE) and the allele-frequency (af/AF). If the object is not provided the tool assumes there are no somatic small variants.

germSmallVars	GRanges object containing all germline small variants (SNV and INDEL). Required metadata columns are reference base (ref/REF), alternative base (alt/ALT), annotation of the gene name (gene/GENE) and the allele-frequency (af/AF) If the object is not provided the tool assumes there are no germline small variants.
ploidy	ploidy of the sample (numeric value)
colnameTcn	character indicating the name of the metadata containing the tcn information in the somCna object. If not provided the tool tries to detect the column according to default names
colnameCnaType	character indicating the name of the metadata containing cna type information in the somCna object. If not provided the tool tries to detect the column according to default names
includeHomoDel	default = TRUE; if FALSE homozygous deletions are excluded
includeIncompleteDel	default = TRUE; if FALSE heterozygous deletions are excluded
assumeSomCnaGaps	(logical, default=FALSE) Only required if the somCna object lacks copy number information for genomic segments on which small variants are detected. By default, variants in such regions will be excluded from the analysis as required information about the copy number is missing. These variants will be attached to the final output list in a separate tibble. To include them, this flag must be set TRUE and the ground ploidy must be given as an input. This ground ploidy will then be taken as tcn in the missing regions. If no ploidy is given the tool will assume the ground ploidy of 2 when this flag is TRUE.
byTcn	logical, default=TRUE; optional if includeHomoDel or includeIncompleteDel is TRUE. If FALSE the tool will not use tcn as a criterion to assign large deletions. It will use the cna_type column and check for indicating strings like HOMDEL/HomoDel/DEL. Some commonly used strings are covered. It is recommended to leave this flag TRUE
ZP_env	internal variable... not recommended to be changed by user
verbose	logical, default=FALSE; prints functions that are called

Value

A list containing tibbles with all input variants

Examples

```
cnvs = GenomicRanges::GRanges(
  dplyr::tibble(
    chr = "chr17",
    start = c(170060, 34520990),
    end = c(34520990, 83198614),
    tcn = c(2, 1),
    cna_type = c("neutral", "LOH")
  )
)
somatic_vars = GenomicRanges::GRanges(
```

```

dplyr::tibble(
  chr="chr17",
  start = 7675088,
  end = 7675088,
  ref = "C",
  alt = "T",
  af = 0.65,
  gene = "TP53"
)
)
germline_vars = GenomicRanges::GRanges(
  dplyr::tibble(
    chr="chr17",
    start = 41771694,
    end = 41771694,
    ref = "GTGT",
    alt = "G",
    af = 0.95,
    gene = "JUP"
  )
)
reference = GenomicRanges::GRanges(
  dplyr::tibble(
    chr = "chr17",
    start = c(7661778, 41754603),
    end = c(7687538, 41786931),
    gene = c("TP53", "JUP")
  )
)
)
sex = "female"
purity = 0.9
predict_per_variant(purity = purity, sex = sex,
  somCna = cnvs,
  somSmallVars = somatic_vars,
  germSmallVars = germline_vars,
  geneModel = reference
)

```

predict_zygosity

predicts zygosity of a set of genes of a sample

Description

predicts zygosity of a set of genes of a sample

Usage

```

predict_zygosity(
  purity,
  sex,

```

```

    somCna,
    geneModel,
    bamDna,
    somSmallVars = NULL,
    germSmallVars = NULL,
    bamRna = NULL,
    ploidy = NULL,
    colNameTcn = NULL,
    colNameCnaType = NULL,
    includeHomoDel = TRUE,
    includeIncompleteDel = TRUE,
    showReadDetail = FALSE,
    printLog = FALSE,
    assumeSomCnaGaps = FALSE,
    byTcn = TRUE,
    vcf = NULL,
    haploBlocks = NULL,
    distCutOff = 5000,
    verbose = FALSE,
    debug = FALSE,
    logDir = NULL,
    snpQualityCutOff = 1,
    phasingMode = "fast",
    AllelicImbalancePhasing = FALSE
)

```

Arguments

purity	purity of the sample (numeric value between 0 and 1 indicating the fraction of relevant sample with control/unrelevant tissue)
sex	sex of the sample (character: "male", "female", "m", "f")
somCna	GRanges object containing all genomic regions with annotated total copynumber and cna_type as metadata columns. The total-copynumber column should be named "tcn" but also some other commonly used names. It should contain numeric values or characters that can be converted to numeric values. The cna_type column must contain the information about loss of heterozygosity (LOH). Therefore the term "LOH" must be explicitly mentioned in the column. If a genomic region is not present in the object, it will be taken as heterozygous with neutral TCN of 2.
geneModel	GRanges object containing the gene-annotation of the used reference genome with metadata column of the gene name (gene)
bamDna	path to bam-file
somSmallVars	GRanges object containing all somatic small variants (SNV and INDEL). Required metadata columns are reference base (ref/REF), alternative base (alt/ALT), annotation of the gene name (gene/GENE) and the allele-frequency (af/AF). If the object is not provided the tool assumes there are no somatic small variants.

germSmallVars	GRanges object containing all germline small variants (SNV and INDEL). Required metadata columns are reference base (ref/REF), alternative base (alt/ALT), annotation of the gene name (gene/GENE) and the allele-frequency (af/AF) If the object is not provided the tool assumes there are no germline small variants.
bamRna	optional; path to rna file (bam format)
ploidy	ploidy of the sample (numeric value)
colnameTcn	character indicating the name of the metadata containing the tcn information in the somCna object. If not provided the tool tries to detect the column according to default names
colnameCnaType	character indicating the name of the metadata containing cna type information in the somCna object. If not provided the tool tries to detect the column according to default names
includeHomoDel	default = TRUE; if FALSE homozygous deletions are excluded
includeIncompleteDel	default = TRUE; if FALSE heterozygous deletions are excluded
showReadDetail	default = FALSE; if TRUE a table is added to the output, containing all used reads/rea-pairs with annotated read classification (mut1, mut2, both, none, skipped, dev_var)
printLog	default = FALSE; if TRUE the gene which is evaluated is printed in console, containing the query-name of each read which was used to perform haplotype-phasing and the info into which class it was assigned.
assumeSomCnaGaps	(logical, default=FALSE) Only required if the somCna object lacks copy number information for genomic segments on which small variants are detected. By default, variants in such regions will be excluded from the analysis as required information about the copy number is missing. These variants will be attached to the final output list in a separate tibble. To include them, this flag must be set TRUE and the ground ploidy must be given as an input. This ground ploidy will then be taken as tcn in the missing regions. If no ploidy is given the tool will assume the ground ploidy of 2 when this flag is TRUE.
byTcn	logical, default=TRUE; optional if includeHomoDel or includeIncompleteDel is TRUE. If FALSE the tool will not use tcn as a criterion to assign large deletions. It will use the cna_type column and check for indicating strings like HOMDEL/HomoDel/DEL. Some commonly used strings are covered. It is recommended to leave this flag TRUE
vcf	character; path to variant call file (.vcf.gz format). Will be used (if provided) for extended SNP phasing if variants on the same gene are too far away from each other for direct haplotype phasing
haploBlocks	GRanges object containing haploblocks. Haploblocks are defined as genomic regions in which SNPs are phased to a specific allele. For example a haploblock could be chr1:1000-10000. This would mean that every genotype annotation in the format "1 0" or "0 1" of a SNP in this region will be used to phase somatic variants and define their genotype
distCutoff	numeric, default=5000; if input vcf is provided and SNP phasing is performed, this will limit the distance at which the SNP phasing should not be tried anymore.

As the probability of finding overlapping reads at such a long distance is very low and the runtime will increase exponentially.

verbose	logical, default=FALSE; prints functions that are called
debug	logical, default=FALSE; prints output for debugging
logDir	character; path to directory where logfiles and detailed infos of the run can be stored, if not given, no details will be stored or printed
snpQualityCutOff	numeric, default=1; Cutoff to filter for SNPS that can be used for phasing
phasingMode	character, default="fast"; if set to full. Even if high confidence phasing result could be achieved, following phasing approaches will be carried out
AllelicImbalancePhasing	logical, default=FALSE. Enables allelic imbalance phasing if TRUE

Value

A list of dataframes. Those are the evaluation per variant, the evaluation per gene and, if performed, the info about the haplotype-phasing.

Examples

```
cnvs = GenomicRanges::GRanges(
  dplyr::tibble(
    chr = "chr17",
    start = c(170060, 34520990),
    end = c(34520990, 83198614),
    tcn = c(2, 1),
    cna_type = c("neutral", "LOH")
  )
)
somatic_vars = GenomicRanges::GRanges(
  dplyr::tibble(
    chr="chr17",
    start = 7675088,
    end = 7675088,
    ref = "C",
    alt = "T",
    af = 0.65,
    gene = "TP53"
  )
)
germline_vars = GenomicRanges::GRanges(
  dplyr::tibble(
    chr="chr17",
    start = 41771694,
    end = 41771694,
    ref = "GTGT",
    alt = "G",
    af = 0.95,
    gene = "JUP"
  )
)
```

```

)
reference = GenomicRanges::GRanges(
  dplyr::tibble(
    chr = "chr17",
    start = c(7661778, 41754603),
    end = c(7687538, 41786931),
    gene = c("TP53", "JUP")
  )
)
sex = "female"
purity = 0.9
bamfile <- system.file("extdata", "ZP_example.bam",
  package = "ZygoistyPredictor")
predict_zygoisty(purity = purity, sex = sex,
  somCna = cnvs,
  somSmallVars = somatic_vars,
  germSmallVars = germline_vars,
  geneModel = reference,
  bamDna = bamfile
)

```

ZP_ov

accessor for ZygoistyPredictor runs. Prints an overview about the run

Description

accessor for ZygoistyPredictor runs. Prints an overview about the run

Usage

```
ZP_ov(fp)
```

Arguments

fp full prediction (output of predict_zygoisty())

Value

prints overview about run from function predict_zygoisty()

Examples

```

cnvs = GenomicRanges::GRanges(
  dplyr::tibble(
    chr = "chr17",
    start = c(170060, 34520990),
    end = c(34520990, 83198614),
    tcn = c(2, 1),
    cna_type = c("neutral", "LOH")
  )
)

```

```
)
somatic_vars = GenomicRanges::GRanges(
  dplyr::tibble(
    chr="chr17",
    start = 7675088,
    end = 7675088,
    ref = "C",
    alt = "T",
    af = 0.65,
    gene = "TP53"
  )
)
germline_vars = GenomicRanges::GRanges(
  dplyr::tibble(
    chr="chr17",
    start = 41771694,
    end = 41771694,
    ref = "GTGT",
    alt = "G",
    af = 0.95,
    gene = "JUP"
  )
)
reference = GenomicRanges::GRanges(
  dplyr::tibble(
    chr = "chr17",
    start = c(7661778, 41754603),
    end = c(7687538, 41786931),
    gene = c("TP53", "JUP")
  )
)
sex = "female"
purity = 0.9
bamfile <- system.file("extdata", "ZP_example.bam",
  package = "ZygotyPredictor")
fp <- predict_zygoty(purity = purity, sex = sex,
  somCna = cnvs,
  somSmallVars = somatic_vars,
  germSmallVars = germline_vars,
  geneModel = reference,
  bamDna = bamfile
)
ZP_ov(fp)
```

Index

* datasets

- GR_GENE_MODEL, [5](#)
- GR_GERM_SMALL_VARS, [6](#)
- GR_HAPLOBLOCKS, [6](#)
- GR_SCNA, [7](#)
- GR_SOM_SMALL_VARS, [7](#)

[aff_germ_copies](#), [2](#)
[aff_som_copies](#), [3](#)

[gene_ov](#), [4](#)
GR_GENE_MODEL, [5](#)
GR_GERM_SMALL_VARS, [6](#)
GR_HAPLOBLOCKS, [6](#)
GR_SCNA, [7](#)
GR_SOM_SMALL_VARS, [7](#)

[predict_per_variant](#), [8](#)
[predict_zygosity](#), [10](#)

[ZP_ov](#), [14](#)